

A Journal of the Gesellschaft Deutscher Chemiker

Angewandte Chemie

GDCh

International Edition



WILEY-VCH

REPRINT

Reprint

A Journal of the Gesellschaft Deutscher Chemiker

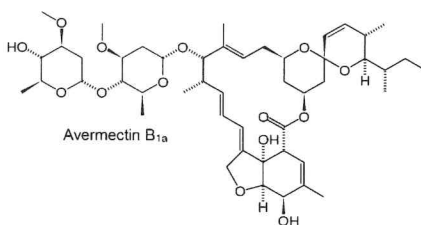
Angewandte Chemie

GDCh

International Edition

Avermectins

A Splendid Gift from the Earth: The Origins and Impact of the Avermectins (Nobel Lecture)



Japanese soil was the origin of one of the most important drugs of the world: ivermectin. No other drug has such importance for the health of millions of people, particularly in the poor regions of the world. The discovery of the parent compounds of the avermectines is described first hand by S. Ōmura.

S. Ōmura* _____ 10190–10209

Keywords: anthelmintics · ivermectin · medicinal chemistry · river blindness · worm infestation

2016 – 55/35

© WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim

WILEY-VCH

Avermectins

International Edition: DOI: 10.1002/anie.201602164

German Edition: DOI: 10.1002/ange.201602164

A Splendid Gift from the Earth: The Origins and Impact of the Avermectins (Nobel Lecture)**

Satoshi Ōmura*

anthelmintics · ivermectin · medicinal chemistry ·
river blindness · worm infestation

Introduction

The origin of one of the world's foremost, revolutionary, versatile yet relatively unknown drugs lies in Japanese soil—literally and metaphorically. Ivermectin, a multipurpose drug derived from a single microscopic organism discovered in Japanese soil, is being taken annually free of charge by over 250 million people—twice as many people as the entire Japanese population. It's impact on improving the overall health and welfare of hundreds of millions of men, women, and children, mostly in poor and impoverished communities, remains unmatched. It continues to defy many preconceived concepts, with no drug resistance developing in humans despite years of extensive monotherapy. This has led to it being included on the World Health Organization's "List of Essential Medicines", a compilation of the most important medications needed in any basic health system. Several international public health experts have also taken the unprecedented step of recommending mass administration of ivermectin to all members of often polyparasitized communities in developing countries as a simple, prophylactic and curative public health intervention.^[1]

Ivermectin, along with its parent compound, avermectin, are both extremely broad-spectrum antiparasitic agents. Ivermectin is among those few compounds, such as penicillin and aspirin, delineated as "wonder drugs", all, incidentally, originating from natural products. It is also predominantly a drug for the poor. It is being increasingly used to eliminate intractable tropical diseases, as well to tackle an ever-increasing range of diseases, and showing promise to provide a solution to hitherto indomitable public health problems. It remains the most potent anti-infective agent in clinical use; the safe single adult dose of around 12 mg once a year comparing favorably with antibiotics such as penicillin and tetracycline that require doses of 1000 mg or more per day.

The avermectins, and the derivative ivermectin, were identified in the mid-1970s. The discovery was exceptional, as avermectin represented the world's first "endectocide", a term specially created to describe the compound, which was capable of killing a wide variety of parasitic and health-threatening organisms both inside and outside the body. The avermectins were found to be two- to threefold more potent than compounds in use at the time. Moreover, ivermectin was found to be effective orally, topically, or parentally and showed no signs of cross-resistance with commonly used antiparasitic agents.^[2-4] Since its discovery, the benefits of

ivermectin in terms of global public health and socioeconomic welfare (direct and indirect) have been immeasurable and they continue to accumulate. The discovery occurred at a time when the international community was focusing attention on disregarded and seemingly unconquerable diseases which had been plaguing resource-poor populations throughout the tropics for centuries. The advent of the antiparasitic ivermectin provided a safe, simple, and effective solution, now driving several of those seemingly invincible tropical diseases to the brink of eradication.

In science, as elsewhere, it is individuals who are the true agents of change. The discovery of the avermectins was the result of a novel international multidisciplinary research project between a public sector institution (Japan's Kitasato Institute) and a private sector pharmaceutical company (the US-based Merck, Sharp & Dohme). But the successful history of the pioneering public private partnership has been dependent upon the unwavering commitment, ability, and quality of scientific and cultural exchanges among the team of exceptional individuals involved, all of whom managed to overcome differences in nationality, working practices, and sometimes differing goals.

Likewise, the availability of and access to the drug, its distribution, and its enormous and widespread beneficial impact have been dependent upon a combination of an unprecedented drug-donation program plus an exceptional, ground-breaking, multifaceted international partnership incorporating, among others, the public and private sectors, multilateral agencies, donor organizations, governments, non-governmental organizations, scientists, health workers, and entire disease-affected communities.

Ivermectin: Preparing the Ground

In Japan, the Kitasato Institute (KI), founded in 1914 by Shibasaburo Kitasato—known as the Father of Serotherapy and nominated for the first Nobel Prize in 1901—has long been recognized as a world-leading center for the discovery of

[*] Prof. Dr. S. Ōmura
Kitasato University, Kitasato Institute for Life Sciences
Minato-ku, 9-1, Shirokane 5-chome, Tokyo, 108-8642 (Japan)
E-mail: omuras@insti.kitasato-u.ac.jp

[**] Copyright© The Nobel Foundation 2015. We thank the Nobel Foundation, Stockholm, for permission to print this lecture.

drugs and vaccines, primarily those derived from natural sources. Investigative research and development of chemotherapeutic drugs for practical use is a fundamental core of the work of the institute. Over 100 years ago, in a pioneering breakthrough, Sahachiro Hata, working with Paul Ehrlich, developed salvarsan, a remedy for syphilis, which was a major global health problem at the time. Salvarsan is widely recognized as the world's first chemotherapeutic drug. In the 1930s, Zenjiro Kitasato's research into plant alkaloids and terpenoids later led to the development of the antitussive compound sapogenin. In the late-1940s, Toju Hata conducted research on antibiotics produced by microbes, which led to the discovery of leucomycin in 1953 and the anticancer compound mitomycin in 1956.

In the mid-1960s, from a background of studying aspects of fermentation and having garnered significant experience in using the, at the time, novel and nascent nuclear magnetic resonance (NMR) spectroscopy to determine the structure of organic compounds, I was fortunate to join the institute's illustrious alumni, which also includes Kiyoshi Shiga and Hideyo Noguchi.

Shortly after joining the KI, having worked on identifying the stereostructure of a handful of compounds, I realised that I could only identify hard-to-find compounds that others had spent a great deal of time, effort, and expertise in discovering. Consequently, I decided to challenge myself to actually undertake the discovery process, which was fundamental to identifying new compounds and microbial metabolites, as well as investigate their structure and possible bioactive properties. To that end, coming from a farming family background, through which I had developed a profound respect for nature and its role as a primary source of most of the materials we need for survival, I opted to concentrate on soil microorganisms. Soils often contain 10^9 to 10^{10} microorganisms per gram (dry weight), which possibly represents in excess of 1 million bacterial species^[5] and, in my experience, around one third of soil samples tested produce antimicrobial substances. Unfortunately, there is no accepted "Gold Standard" method for isolating and identifying soil bacteria or other microorganisms. A serial dilution and spread-plate method is a reasonably good starting point for isolating bacterial colonies from soil, but even at this early stage the choice of isolation medium is critical and depends on the specific goals. Consequently, devising mechanisms to cope sensibly with this enormous diversity is essential. As a result, I refocused my research on the search for new antibiotics and other biologically interesting microbial metabolites, such as growth factors, enzymes, and enzyme inhibitors, based on my conviction that new and innovative screening systems were the key to discovering new compounds—a belief that I have steadfastly maintained to this day.

In science, knowledge and understanding no longer appear quickly. Time, patience, trial, and error are all essential ingredients in any screening process. Most screening systems retain their effectiveness but, over the years, I have devised and implemented one or two new screening mechanisms annually, discarding existing systems when resources did not permit them to be kept in operation. Generally, we

now routinely have at least 10 customized screening systems operational.

Although many screens prove successful, others do not yield the results envisaged, although this does not mean they are nonfunctional. In this matter, I have always been guided by the words of Louis Pasteur: "Chance favors the prepared mind". I believe that this is the key to investigating and unravelling the mysterious world and secrets of microorganisms. This is the mindset that I have always followed and which has allowed nature to reveal to me almost 500 microbial metabolites that have unique or useful bioactive properties, several of which have proved of incalculable benefit, direct and indirect, to humankind (Figure 1).

• Microorganisms:	
New genera	13
New species & sub-species	53
• New compounds	483
• Useful compounds	26
• Targets for total syntheses	>100

Figure 1. Discoveries (1965–2014).

The painstaking work at the KI involves many of the first steps on a long road to the creation of a successful drug or useful chemical reagent. We take samples from nature that contain microorganisms. We then allow the microbes in the sample to grow on agar media plates, slowly cultivating them to produce a pure cultivar, making sure that we concentrate on novel types. The organism and strain are then identified, grown in liquid culture, and a culture broth is formed. We carry out initial assays on the broth, including an initial metabolite analysis. In the case of the microbe that was the origin of ivermectin, for example, we identified that it also produced a toxic compound, oligomycin, knowledge that proved to be of great value with respect to explaining toxicity problems during later tests in animal models. Once these initial steps have been completed, we can scale-up using a jar fermenter, which facilitates clearer identification of the organism and its preservation as well as purification and structural analysis of any promising compound (Figure 2). We then conserve all the microorganisms and compounds in our libraries for future testing and evaluation, either by KI scientists or others.

Generally, during our routine discovery work, we deliberately select unusual microorganisms with the intent to maximize the chances of finding new compounds. In addition, we generally do not have a single, specific objective, preferring to apply initial screens for a variety of bioactive properties. The characteristics of the microbe that we isolated and cultured at the Kitasato Institute, and which produced the avermectins, were unique and were critical elements in the discovery process.^[6]

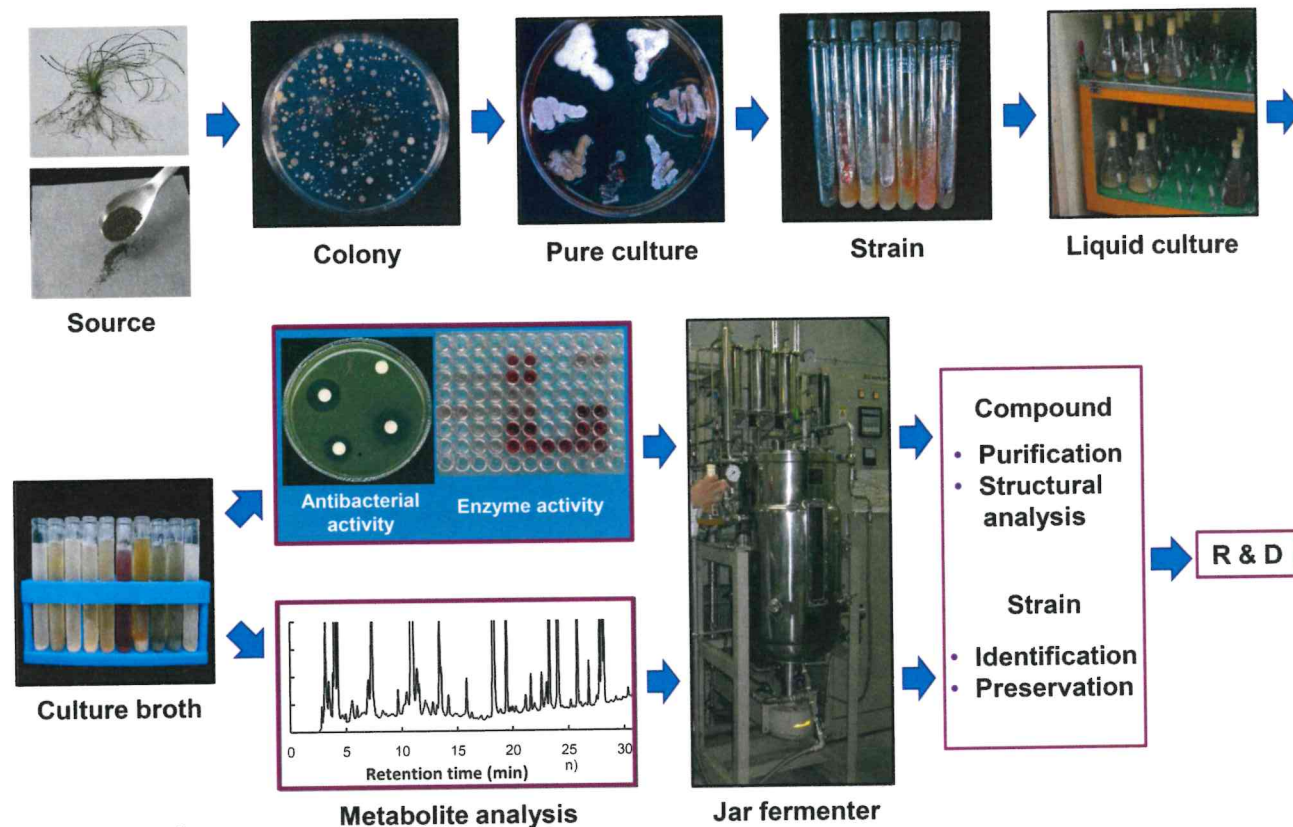


Figure 2. Screening for new bioactive compounds.

From the outset of my research I determined that it was highly useful to identify not just a new compound but also the microbe that produced it, usually placing both together in a visual presentation—a traditional that I shall maintain in this Review. We have attempted to isolate microbes from every kind of natural environment, primarily from soil and latterly from seaweed, plant leaves, and plant roots. Figure 3 shows the diversity of microorganisms that have been isolated from plants roots compared to soil and provides an indication of how the source can significantly impact the type of microbe found. For example, we have recently identified two new compounds, spoxazomycin (Figure 4),^[7] which displays anti-trypanosomal activity, and trehangelin (Figure 5),^[8] a photo-oxidative hemolysis inhibitor from plant-root origins.

It goes without saying that all microbes and chemicals are small, well beyond human visual acuity. It therefore seemed sensible to find mechanisms that would clearly signal the presence of something new or potentially useful. Mindful of the fact that, throughout human history, alkaloids, mostly from plant sources, have been the mainstay of traditional medicine, I decided to introduce a new method of chemical screening. This entailed a search and isolation method to identify organic compounds in fermentation broths by employing a simple color-change reaction. I decided to utilize a simple mechanism using Dragendorff's reagent. Alkaloids, if present, react with the reagent—which contains bismuth nitrate and potassium iodide—to produce an easily visible orange or orange-red precipitate.

We implemented this screening system in 1968, based on my profound belief that microorganisms never engage in futility, it is just our lack of knowledge and vision that prevents us from understanding what they produce, how, and for what purpose. The first compound isolated through this chemical screening system was the antimicrobial pyridicin (Figure 6).^[9] Of far greater significance, in 1977 we isolated the world's first naturally occurring indolocarbazole compound, staurosporine, produced by the bacterium *Streptomyces staurosporeus* (Figure 7).^[10,11] Nine years later, Dr. T. Tamaoki found that staurosporine possessed the ability to potently inhibit the functioning of protein kinase C (PKC), the first such compound identified to do so. PKCs are a family of enzymes that cause increased expression of oncogenes, thereby promoting cancer progression.^[12] Almost immediately, staurosporine became one of the world's most prominent research reagents of microbial origin and proved to be the forerunner of many of the recently introduced anticancer agents. For example, the development of imatinib (Gleevec; Figure 8) has been directed and influenced by the unique chemical structure and biological activity of staurosporine.^[13] For me, the discovery of staurosporine was a significant milestone, not just because of its major impact on science and biomedicine, but because it was a vindication of my beliefs that microorganisms offer a virtually unlimited panoply of beneficial products; it is simply a matter of us finding ways to identify and apply them for the good of human society. I also firmly believe that the work that I accomplish and the

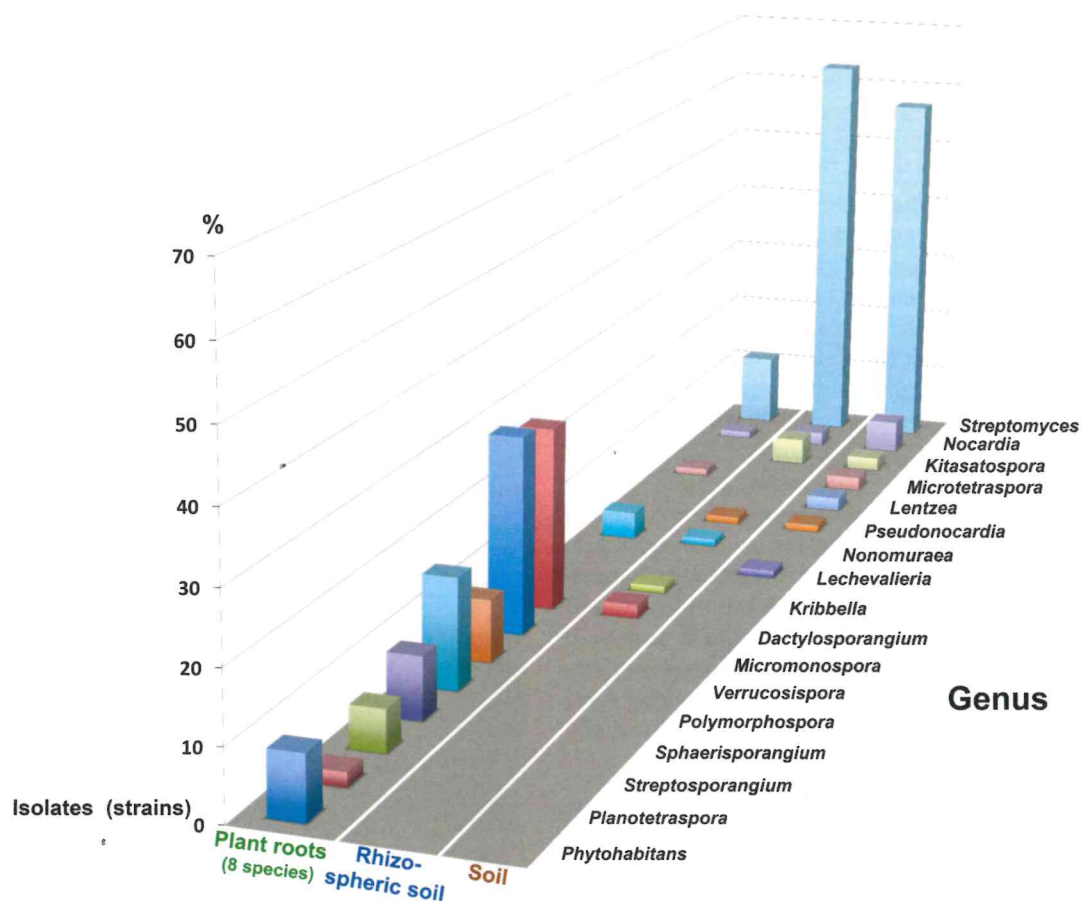
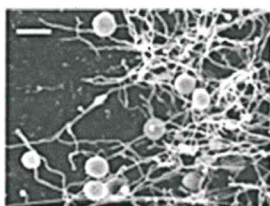


Figure 3. Actinomycetes (from plant root and soil samples).



Streptosporangium oxazolinicum K07-0460^T
(Bar: 10 μM)

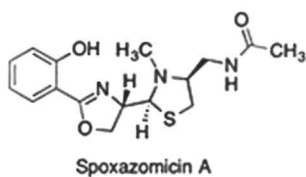
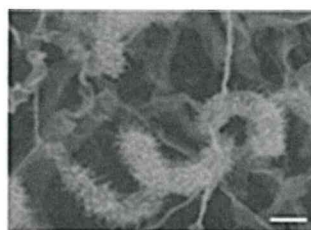


Figure 4. Spoxazomicin A.



Streptomyces griseoflavus subsp.
pyridicus NA-15^T (Bar: 1 μM)

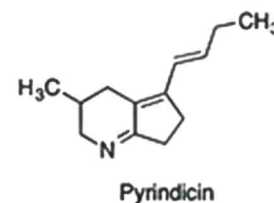
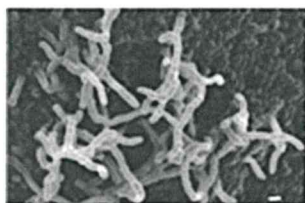


Figure 6. Pyridicin.



Polymorphospora rubra K07-0510
(Bar: 1 μM)

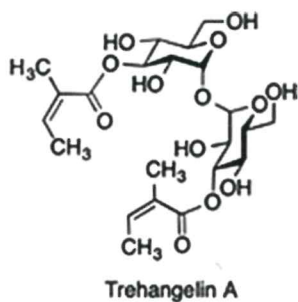
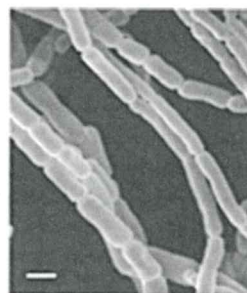


Figure 5. Trehangelin A.



Saccharothrix aerocolonigenes subsp.
staurosporeus AM-2282^T
(*Lentzea albida* AM-2282)
(Bar: 1 μM)

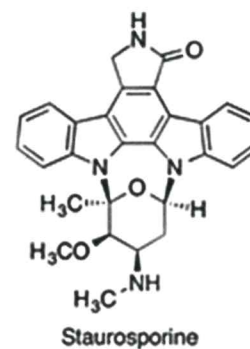
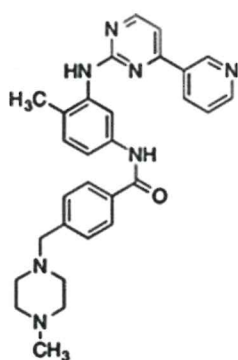


Figure 7. Staurosporine.

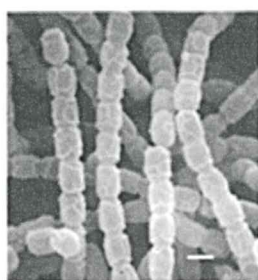


Imatinib (Gleevec®)

Figure 8. Imatinib.

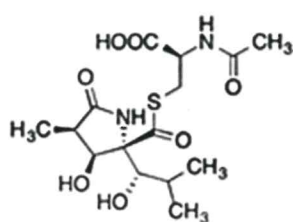
compounds identified and stored can be taken forward or exploited by others for the good of all.

Another novel screening system led to the discovery of lactacystin (Figure 9), an inhibitor of proteasomes. Lactacystin was found by a method involving induction of neurite outgrowths in Neuro2a, a cell line of murine neuroblastoma cells.^[14] This compound proved to be the forerunner for the anticancer agent bortezomib (Velcade; Figure 10).

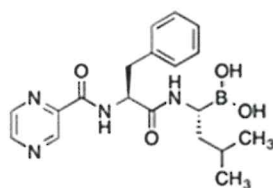


Streptomyces lactacystinicus OM-6519T
(Bar: 1 μM)

Figure 9. Lactacystin.



Lactacystin



Bortezomib (Velcade®)

Figure 10. Bortezomib.

The experience, techniques, and knowledge gained at the KI in isolating microorganisms, cultivating them, identifying them, and then determining the compounds they produce, analyzing the chemical structure, and elucidating their biological or chemical properties provided an optimal basis for the discovery of ivermectin. However, although we possessed the skill and expertise to discover novel microorganisms and chemicals, we had neither the techniques nor the resources to carry out the requisite research and development essential for taking a promising compound through the extremely expensive and often disappointing drug production pipeline. To accomplish that task requires the commitment and extensive resources of a major commercial partner.

Ivermectin: The Beginnings

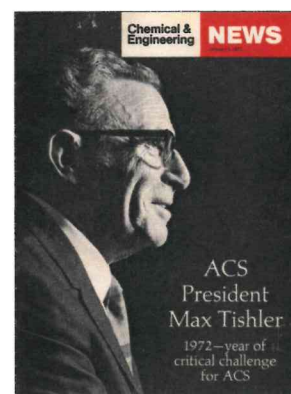
In the early-1970s, Prof. Yukimasa Yagisawa, General Manager of the Japan Antibiotics Research Association (JARA), encouraged me to exploit the possibilities of research work overseas and the benefits it could provide for both myself and for Japan. He kindly introduced me to key individuals in his network of overseas connections and, as a consequence, in 1971 I was granted a sabbatical that allowed me to take up an invitation from Prof. Max Tishler to work as Visiting Research Professor in his newly formed Chemistry department at Wesleyan University (Figure 11). Max, who



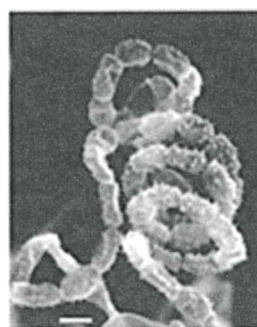
Satoshi Omura Max Tishler (1906-1989)

Figure 11. Ivermectin: The beginning.

Wesleyan University
USA (1972)

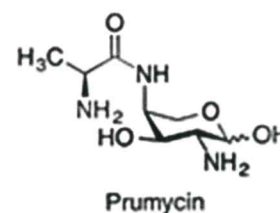


almost immediately became President of the American Chemical Society (ACS), had established the department following retirement from his position as President of the MSD Research Laboratory (MDRSL), where he had had a long and distinguished career. My initial work in his laboratory focused on the structural analysis of a new antibiotic, prumycin (Figure 12)^[15] that I had discovered prior to my departure from Japan, as well as on the structure/activity relationships of macrolides^[16] and the mode of action of cerulenin (Figure 13). The contribution that both of these individuals made to my development as a scientist, educator, and individual has been inspirational and beyond measure.



Streptomyces kagawaensis F-1028T
(Bar: 1 μM)

Figure 12. Prumycin.



Prumycin

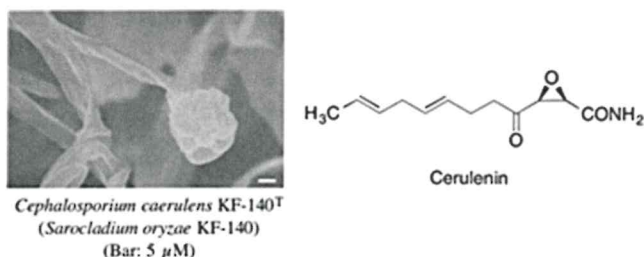


Figure 13. Cerulenin.

My intended stay in the US was curtailed, as I was recalled to head the Research Department at the KI, following the retirement of the then director, and I returned in early 1973. In view of my impending return, and extremely mindful of the critical need to obtain funds to support research work in Tokyo after I returned, I visited many major US pharmaceutical companies, presenting a proposal for a collaborative research project. I was greatly encouraged because, as I had previously discovered several antibiotics, such as the aforementioned prumycin (an antifungal agent)^[17] and cerulenin (an antifungal and inhibitor of fatty acid biosynthesis),^[18] as well as leucomycin A3 (an antimicrobial; Figure 14),^[19] all of

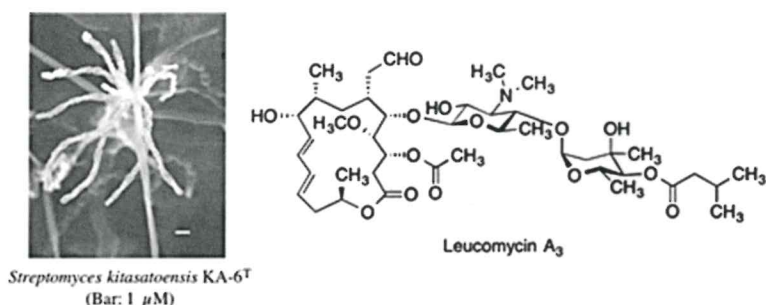


Figure 14. Leucomycin A3.

the companies were supportive. At the time, Max, who knew my work and ideas very well, discussed my plan with Dr. L. H. Sarett, Max's successor at MSDRL, with whom he had worked closely for many years. Max's close connection with Merck and his personal linkage to Dr. Lew Sarett expedited a research collaboration with the MSDRL, which commenced in April 1973. Individuals who played key roles in the alliance are shown in Figure 15. Initially, the goal was to find growth-promoting antibiotics suitable for animals, enzyme inhibitors, and general purpose antibiotics produced by microorganisms, but the work soon expanded to encompass other targets.

Ivermectin: The Advent and Use in Animals

As the basis of the research initiative, the KI carried out the isolation of what we identified as extraordinary microorganisms, culturing them, and then undertaking preliminary in vitro evaluation of the bioactivity of any compounds we deemed to be of potential interest, prior to sending the most

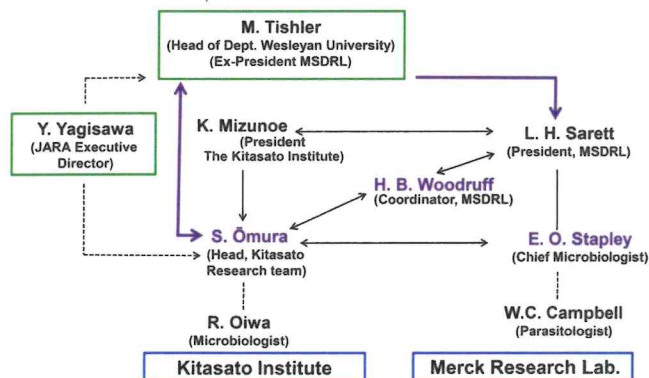


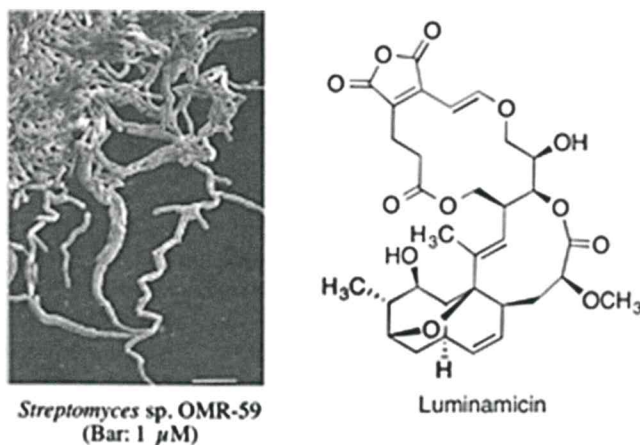
Figure 15. The Kitasato-MSDRL collaboration (1973).

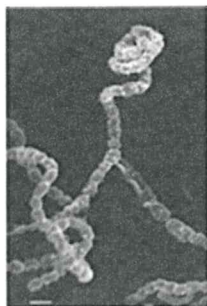
promising—from our existing library and from newly identified specimens—to MSDRL for in vivo testing.

As a result of the collaboration, a variety of compounds were discovered, the majority exhibiting a range of interesting biological activities and structures. These included lumnamycin (Figure 16),^[20] an anti-anaerobic bacterial, vineomycin A1 (Figure 17)^[21] and setamycin (Figure 18),^[22] both of which have unique structures, elasnin (Figure 19),^[23] the first human elastase inhibitor of microbial origin, and factumycin (Figure 20), a growth-promoting antibiotic for veterinary use.^[24]

Of far greater import was avermectin. Simply put, avermectin proved to be one of the world's most remarkable biomedical discoveries, being accompanied by a number of world "firsts" and having immeasurably beneficial impact on animal and human health worldwide.

As part of their new in vivo evaluation, and following a suggestion from Max Tishler, the MSDRL introduced a new program to screen fermentation broths that we identified as being promising.^[25] This was done because there was confidence that our broths likely contained interesting compounds. In addition, adding a fermentation broth to the feed of a single animal means it can be tested simulta-

Figure 16. Luminamicin. Scale bar: 1 μ m.



Streptomyces matensis subsp.
vineus OS-4742^T (Bar: 1 μM)

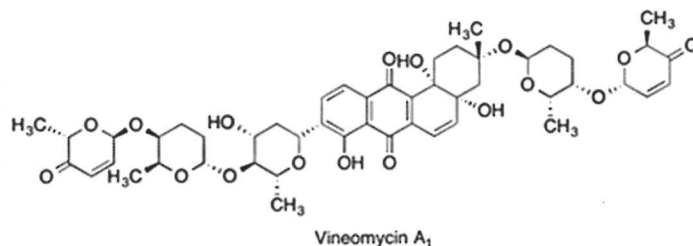
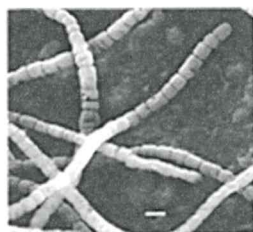


Figure 17. Vineomycin A₁.



Kitasatospora setae KM-6054^T
(Bar: 1 μM)

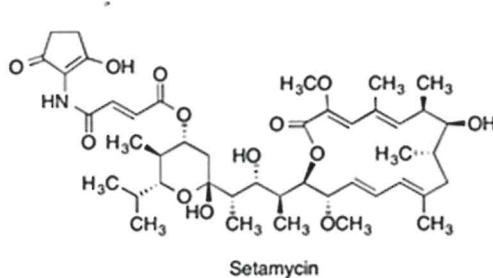
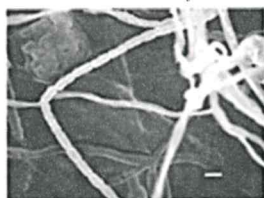


Figure 18. Setamycin.



Streptomyces noboritoensis KM-2753
(Bar: 1 μM)

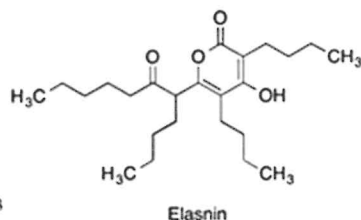
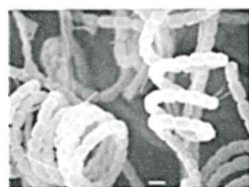


Figure 19. Elasin.



Streptomyces lavendulae OS-4369
(Bar: 1 μM)

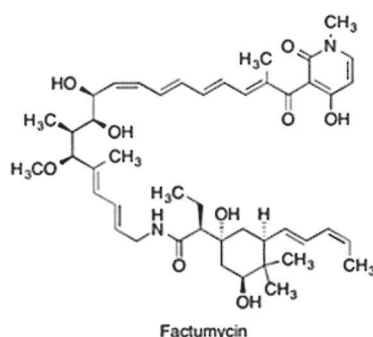


Figure 20. Factumycin.

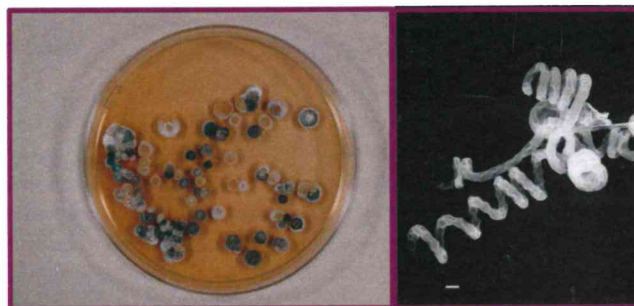
neously for both efficacy and toxicity, often with results appearing in a week rather than the weeks or months that are needed using *in vitro* tests.

MSDRL researchers screened our microorganisms, which were produced according to our description of the necessary fermentation conditions, the fermentation broths being tested in a novel model of helminth infection in which mice were

infected with the nematode worm *Nematospiroides dubius*.^[26,27] In one of the first 50 specially selected microorganisms we sent in 1974, Dr. William Campbell and his team found an actinomycete, strain MA-4680, which produced a compound possessing excellent antihelminthic activity with little or no toxicity. The unpurified broth killed all the intestinal worms and removed all signs of parasite eggs from the animal's faeces.

The producing microorganism (Figure 21) was originally named *Streptomyces avermitilis* MA-4680 but, in 2002, based on characterization of the original strain and morphological and phylogenetic comparisons, including 16S rDNA sequencing, with closely related members of the genus *Streptomyces*, it was proposed that the organism was in fact a new species and renamed *Streptomyces avermectinarius*.^[28]

After a few trials to confirm the bioactivity findings, isolation chemists were engaged to identify the causal entity. The active ingredient of the

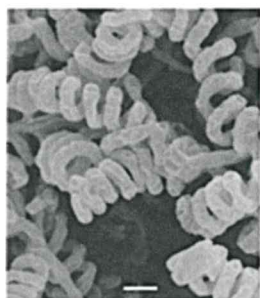


Streptomyces avermectinarius (*S. avermitilis*)

Figure 21. The avermectin-producing strain. White scale bar: 1 μm.

broth was identified and named avermectin, which MSDRL chemists found to be a complex mixture of 16-membered macrocyclic lactones, with the fermentation of *S. avermectinarius* producing a mixture of eight avermectin compounds (A1a, A1b, A2a, A2b, B1a, B1b, B2a, and B2b) (Figure 22). Compounds of the B series containing a 5-hydroxy group are markedly more active than those of the A series, which contain a 5-methoxy group. The four main components, avermectin A1a, A2a, B1a, and B2a, constituted 80% of the mixture, with the rest composed of four lower homologues A1b, A2b, B1b, and B2b. The structure of the compound was also swiftly elucidated, and it was fast-tracked for development.^[29]

In 1979, the first papers on the avermectins were published, describing the chemicals as a series of macrocyclic lactone derivatives possessing extraordinarily potent anthelmintic properties.^[30–32] Up until that time, only a handful of



Streptomyces avermectinius
(*S. avermitilis*) MA-4680^T
(Bar: 2 μM)

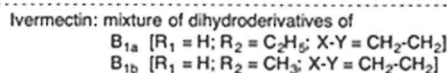
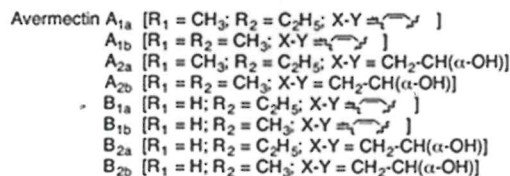
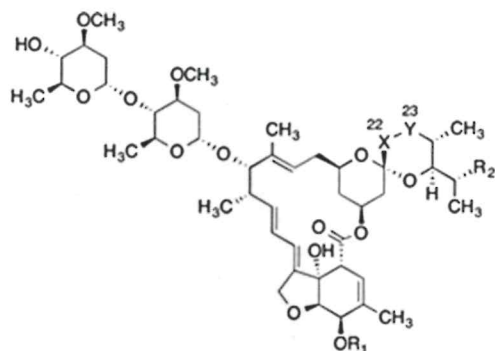


Figure 22. Avermectin.

the several thousand microbial fermentation products discovered exhibited any anthelmintic characteristics. Although structurally similar to macrolide antibiotics and antifungal macrocyclic polyenes, the avermectins did not demonstrate any antibacterial or antifungal activities.

An interdisciplinary team at MSDRL, headed by William Campbell, further investigated the eight active compounds, of which avermectins B_{1a} and B_{1b} were found to have the highest activity. Reduction of the C₂₂–C₂₃ double bond of B_{1a} and B_{1b} compounds with the Wilkinson catalyst improved both the spectrum of activity and safety, and the resulting 22,23-dihydro-B₁ complex (as a mixture of 80% B_{1a} and 20% B_{1b}) was selected for further commercial development under the generic, nonproprietary name, ivermectin.^[33]

The avermectins proved to be effective against roundworms of the intestinal and respiratory tracts as well as filarial parasites^[34] and demonstrated biocidal activity against a diverse range of nematodes, insects, and arachnids. The mode of action turned out to be both unique and robust, and was 25 times more potent than all currently available anthelmintics. Further analysis revealed that ivermectin was highly efficacious against mite, tick, and botfly ectoparasites, organisms that cause massive economic losses in the livestock industry. MSDRL researchers also observed that the compound had remarkable activity against external and internal parasites in horses, cattle, pigs, and sheep, being effective against, among others, gastrointestinal roundworms, lungworms, mites, lice, and hornflies. It was also found to be successful in treating larval heartworms in dogs, but not adult worms, and could be used to treat mange and other conditions in canines. However, no activity was found against flatworms, protozoa, bacteria, or fungi.^[35–38]

The avermectins broad spectrum of activity, wide therapeutic index, and novel mode of action resulted in them being introduced onto the Animal Health market in 1981. Two years after their introduction, avermectin-derivative

products became the international veterinary sector's biggest seller, accruing annual sales income of around \$1 billion, a position maintained for a quarter of a century, and the ivermectin-based parasiticide products reportedly becoming the company's fifth best-selling product group.^[39]

MSDRL research staff and others around the world have exhaustively searched since the original discovery, but no other avermectin-producing organism has ever been found. The strain that we isolated from a single soil sample collected near a golf course bordering the ocean at Kawana in Ito City in the Shizuoka region of Japan remains the only avermectin-producing organism ever found.

Dr. Boyd Woodruff, from MSDRL, was appointed to work alongside our team at the KI in Tokyo, and I am convinced that his personal commitment and expertise were significant factors in making the collaboration such a great success.

Ivermectin: Mode of Action

The avermectins potentiate neurotransmission by boosting the effects of glutamate at invertebrate-specific glutamate-gated chloride channels, with minor effects on γ-aminobutyric acid (GABA) receptors.

In parasites, neurotransmission inhibition occurs via glutamate-gated (Cl⁻) channels in nerve and muscle cells preventing their closure.^[40] This leads to hyperpolarization of the neuronal membrane, inducing paralysis of the somatic muscles, particularly the pharyngeal pump, killing the parasite.^[41,42] GABA-related (Cl⁻) channels are commonplace in nematodes, insects, and ticks.^[43–45] In mammals, GABA receptors and neurons only occur in the central nervous system (CNS) and are thus not accessible,^[46] with ivermectin being safe for vertebrates as it cannot cross the blood–brain barrier. Initial fears that ivermectin was contra-indicated in children under the age of five or who weighed less than 5 kg, where the drug might be able to cross the as yet not fully developed blood–brain barrier, were proven to be unfounded.^[47]

In humans, ivermectin exerts a peculiar and singular effect that remains poorly understood. The immune response to filarial infection is complex, involving Th₂-type systems which counter infective L₃ larvae and microfilariae, whereas a combination of Th₁ and Th₂ pathways are involved in resisting adult worms. It is believed that female adult worms are able to manipulate the immunoregulatory environment to ensure the survival of their microfilarial offspring.^[48] Ivermectin treatment of onchocercal filarial infection causes microfilariae to quickly disappear from the peripheral skin lymphatics. The effect is long-lasting, because adult female worms are prevented from releasing microfilariae.^[49] Dermal microfilarial loads are reduced by 78% within two days, and

by some 98% two weeks after treatment, remaining at extremely low levels for about 12 months. Female worms slowly resume release of microfilaria 3–4 months post-treatment, but at a mere 35% of the original production.^[50] Regular treatment consequently decreases the incidence of infection, interrupts transmission, and reduces morbidity and disability. However, the actual mechanism by which ivermectin exerts its effect on microfilariae remains unclear.^[51]

The half-life of ivermectin in humans is 12–36 h. The lowest levels of dermal microfilariae occur well after this timeframe, meaning that not all microfilariae are killed in the early days, and microfilariae are known to migrate into deeper dermal layers, subcutaneous fat, connective tissue, and lymph nodes following ivermectin administration.^[52] It is now believed that ivermectin somehow prevents microfilariae from evading the immune system, resulting in the host's own immune response killing the immature worms.^[53,54]

Ivermectin does not kill adult worms but suppresses the production of microfilariae by adult female worms, thereby reducing transmission. As the adult worms can continue to produce microfilariae until they die naturally, ivermectin has to be taken once annually for the 16–18 years of the adult worm lifespan in order to break transmission.

Th2 responses instil protective immunity against both L3-infective larvae and the microfilaria stage, but parasites are able to avoid these responses, which may help explain why drug resistance in parasites in humans has not yet appeared.

Ivermectin: Development for Human Use

In the mid-1970s, the global community mobilized itself to address the major problems of neglected tropical diseases. Following the setting up of the Onchocerciasis Control Programme (OCP) in West Africa in 1974, the UN-based Special Programme for Research & Training in Tropical Diseases (TDR) was established in 1975. Onchocerciasis and lymphatic filariasis were two filarial infections among TDR's eight target diseases, with onchocerciasis, at the time, being a major public health problem, affecting 20–40 million people in endemic areas, predominantly in Africa (Figure 23).

Historically found primarily in 30 countries in sub-Saharan tropical Africa, onchocerciasis is caused by a nematode, *Onchocerca volvulus*, which lives for up to 15 years in the human body. Female worms continually produce several millions of microfilaria during their lifetime, with the worms being transmitted to humans via the bite of a blood-feeding blackfly.

At the time, there were no safe and acceptable drugs available to treat onchocerciasis, which had plagued Africa for centuries and nobody was interested in developing anti-onchocerca drugs, as there was no apparent commercial market. Consequently, the OCP based its operations on expensive aerial spraying of insecticides to kill riverine vector fly larvae.

MSDRL scientists soon realised that the anthelmintic potency of ivermectin could help to conquer filarial diseases in humans, and joined forces with the WHO, nongovernmental organizations, international donors, governments, and

- Caused by filarial worms, transmitted by *Simulium* black flies
- Females release millions of immature worms; migrate to skin & eyes - skin disease, unbearable itching & blindness.



• People at risk	120 million
• People infected	18 million
• Blinded / disabled	770,000
• Disease burden (DALY)	1.1 million
• Countries affected	36
• No safe drugs available	(data~1987)

Figure 23. Human health goals: Onchocerciasis (river blindness).

Source: UNDP/World Bank/WHO Special Programme for Research & Training in Tropical Diseases (TDR).

affected communities to drive forward evaluation of the drug.^[55]

Meanwhile, with respect to research needs, TDR identified that the discovery of effective chemotherapeutic agents was the highest priority, with a macrofilaricide (capable of killing adult worms) substantially preferable to a microfilaricide (which would target immature worms).^[56] Research was hampered by the fact that *Onchocerca* species would not develop to maturity in any rodents, making it impossible to screen compounds against the target organism in a suitable animal model. TDR established a tertiary screen, using cattle, for compounds showing positive results in any secondary screen. The screen was the best predictor of what a compound would do in humans, with well over 10 000 compounds being screened.^[57,58]

In reality, ivermectin's role in human medicine began in 1978 inside the MSDRL, with William Campbell being the driving force behind the investigation of the potential for human use. After receiving very positive results after submitting ivermectin to the Australian cattle screen, he subsequently reported to MSDRL management that "an avermectin could become the first means of preventing the blindness associated with onchocerciasis".^[59,60]

In 1981, MSDRL's Mohamed Aziz, previously of the WHO, undertook a small clinical trial of ivermectin in patients, with safety paramount. Commencing with a very low dose of $5 \mu\text{g kg}^{-1}$, he found that a single dose of $30 \mu\text{g kg}^{-1}$ substantially decreased skin microfilariae and confirmed that the effect lasted for at least 6 months, with no serious adverse events. His tests concluded that doses up to $200 \mu\text{g kg}^{-1}$ were safely tolerated.^[61,62]

Ivermectin proved to be ideal for combating onchocerciasis, which has two main manifestations: dermal damage resulting from microfilariae in the skin and ocular damage arising from microfilariae in the eye. Ivermectin proved to slightly increase microfilariae in the eye upon treatment, followed by a gradual reduction, reaching to near zero within six months. This meant little or no ocular damage. The large ivermectin molecule cannot cross the blood/aqueous humour barrier, stopping it entering the anterior chamber and directly

killing or paralyzing microfilariae.^[63] This made ivermectin a perfect intervention for patients with ocular involvement.

Similarly, an evaluation of the impact of ivermectin on dermal microfilariae confirmed that it caused almost complete clearance within two days after treatment, reducing the load to virtually zero within eight days. Ivermectin also produces long-term suppression of circulating microfilariae, making it an ideal treatment for patients with dermal involvement.^[64]

Merck received approval from French authorities in 1987 allowing the human use of ivermectin. In a hitherto unprecedented gesture, immediately following registration, ivermectin (branded as Mectizan) was donated free of charge by Merck & Co. Inc., under the direction of Roy Vagelos (Figure 24), for the treatment of onchocerciasis (river blindness), with KI foregoing all royalties. The donation was for as



The Kitasato Institute (1989)

Figure 24. Ivermectin—The world's most effective drug donation.

long as the drug was required, in the amounts that were needed. This represented the first such large-scale drug-donation initiative and it has resulted in the world's largest, longest-running and most successful drug-donation programme.

Introduced for use in the 11-nation OCP, ivermectin was not a cure. It did not kill adult parasites, with a single annual dose simply suppressing symptom-causing onchocercal microfilaria in the skin and eyes and preventing the disease from progressing.^[65] To prevent transmission, every eligible member of an affected community needed to take the drug. Ivermectin only kills immature worms, so entire communities in disease-endemic areas have to take it for up to 15 years, until the adult female worms die naturally.

Massive clinical trials in Africa proved ivermectin to be a highly effective and safe microfilaricide, which need not be given more frequently than once annually and that has few side effects, which were dose-dependent, mild, and short-lived, with no severe ophthalmological adverse events.^[66–68] Ivermectin is very safe, such that it can be given orally, in the

field, by nonmedical staff, meaning the drug is ideal for mass treatment programs.

The African Programme for Onchocerciasis Control (APOC), established in 1995, built on the success of the OCP and extended community-wide mass drug administration (MDA) of ivermectin to 19 other African nations. APOC is recognized as a cost-effective, large-scale public health intervention of enormous significance, preventing an estimated 17.4 million years worth of healthy life from being lost, and freeing all African children taking ivermectin from the dangers of onchocercal blindness and skin disease.^[69]

In referring to the international efforts to tackle onchocerciasis, in which ivermectin is now the sole control tool, the UNESCO World Science Report concluded, “the progress that has been made in combating the disease represents one of the most triumphant public health campaigns ever waged in the developing world”.^[70]

The success of the campaign to overcome onchocerciasis is due to the sterling efforts and long-term commitment of a truly international, multidisciplinary coalition, some key partners of which are shown in Figure 25.

Key partners for Mass Drug Administration (MDA)

- ✓ Merck & Co. Inc. & Mectizan Donation Program
- ✓ Kitasato Institute
- ✓ World Health Organization (WHO)
- ✓ TDR (Special Programme for Research & Training in Tropical Diseases)
- ✓ Onchocerciasis Control Programme - West Africa (OCP)
- ✓ African Programme for Onchocerciasis Control (APOC)
- ✓ World Bank
- ✓ Endemic country governments
- ✓ Non-Governmental Organizations (NGOs)
- ✓ Affected communities & volunteer drug distributors

Figure 25. Ivermectin distribution.

Effectiveness Against Other Filarial Diseases

Lymphatic filariasis, also known as elephantiasis, is another devastating, highly debilitating disease that threatens over 1 billion people in more than 80 countries (Figure 26). An estimated 120 million people in tropical and subtropical regions are infected, 40 million of whom are seriously incapacitated. The disease results from infection with filarial worms, *Wuchereria bancrofti*, *Brugia malayi*, or *B. timori*. The parasites are transmitted to humans through the bite of an infected mosquito and develop into adult worms in the lymphatic vessels, causing severe damage and swelling (lymphoedema). Adult worms are responsible for the major disease manifestations, the most outwardly visible forms being painful, disfiguring swelling of the legs and genital organs. Around 25 million men have genital disease (most commonly hydrocele) and almost 15 million, mostly women, have lymphoedema or elephantiasis of the leg. The psycho-

- Caused by parasitic worms of the species, *Wuchereria bancrofti* (90%) & *Brugia malayi* (10%), transmitted by various species of mosquitoes



Infection causes filarial fever, elephantiasis, male genital damage & severe social stigma

- People at risk > 1.3 billion
- People infected 120 million
- Countries affected 83
(data ~2000)

Figure 26. Human health goals: Lymphatic filariasis (elephantiasis). Source: Global Alliance to Eliminate Lymphatic Filariasis (GAELF), 2010.

logical and social stigmas associated with the disease are immense, as are the economic and productivity losses it causes.

With respect to the use of ivermectin for lymphatic filariasis, again MSDRL took the initial lead. In the mid-1980s, well before ivermectin was approved for human use to treat onchocerciasis, MSDRL scientists were undertaking trials of ivermectin to measure its impact against lymphatic filariasis and to find optimal treatment dosages.^[71] Meanwhile, TDR was carrying out multicenter field trials in Brazil, China, Haiti, India, Indonesia, Malaysia, Papua New Guinea, Sri Lanka, and Tahiti to evaluate ivermectin, the existing treatment drug DEC, and combinations of the two. The results showed that single-dose ivermectin and single-dose DEC worked as well as each other. The combination, even at low dose, proved even more effective, decreasing microfilarial density by 99% after one year and 96% after two years.^[72–75]

Despite these findings, ivermectin remained unregistered for treatment of lymphatic filariasis until 1998, when approval was granted by French authorities. Several years earlier, another drug, albendazole, produced by SmithKlineBeecham (now GlaxoSmithKline, GSK) had also been shown to be effective in killing both immature and adult worms. Indeed, field trials had confirmed that once-yearly combinations of albendazole plus DEC or ivermectin were 99% effective in ridding the blood of microfilariae for at least a year after treatment. The primary goal of treating affected communities thus became elimination of microfilariae from the blood of infected individuals so that transmission of infection is interrupted. This opened up the prospect of actually eliminating the disease, something that was made eminently possible thanks to GSK agreeing to donate albendazole. In late-1998, following registration of the drug for lymphatic filariasis, Merck extended its ivermectin donation program to cover lymphatic filariasis in areas where it coexisted with onchocerciasis. Subsequently, in 1999/2000, the WHO launched the Global Programme to Eliminate Lymphatic Filariasis (GPELF).

The sheer scale of these disease-elimination enterprises is staggering. During the first decade of this century, some 300 million people, roughly the population of the United

States, were taking ivermectin tablets annually. In 2014, 328 million ivermectin treatments were requested by governments of disease-endemic countries and approved by the Mectizan Donation Committee. Of this total, 73 million were for combined onchocerciasis/lymphatic filariasis treatments, meaning that around 255 million people were due to receive ivermectin treatment during the year (Figure 27). In total,

Ivermectin treatments approved (2014):	
Onchocerciasis	110 million
Lymphatic filariasis	218 million
Sub-total =	328 million
Combined treatments	73 million
TOTAL =	255 million

Ivermectin treatments administered (2013):	
Onchocerciasis	107 million
Lymphatic filariasis	120 million
TOTAL =	227 million

Figure 27. Ivermectin treatments. Source: MDP, WHO(WER), APOC.

1.4 billion ivermectin treatments have been donated for onchocerciasis (1987–2014) and 1.2 billion for lymphatic filariasis (2000–2014). The goal of eliminating onchocerciasis in Latin America by 2015 has virtually been accomplished, with just one endemic area remaining on the border between Brazil and Venezuela in remote Yanomami Indian communities, where some transmission is still occurring.^[76]

Today, despite enormous advances in the fight to conquer onchocerciasis in Africa, and with the elimination target date fast approaching, an estimated 172 million people are still in need of treatment.^[77]

Commercial Ivermectin

Besides donated ivermectin being the sole or primary tool in the two global disease-elimination programs, commercial for-profit preparations of ivermectin-based drugs are also being put to ever-increasing uses. Ivermectin is being used ever more widely as a remedy for strongyloidiasis (which afflicts 30–100 million people worldwide) and to treat and prevent scabies (of which 300 million cases are reported each year). Each year, more uses for the avermectins, and ivermectin in particular, are being found in human and animal health.^[78]

Donated Mectizan is the primary agent for elimination programs for onchocerciasis and lymphatic filariasis (in combination with albendazole). At-cost ivermectin has also now become:

- The drug of choice to treat strongyloidiasis, although it is not available in all nations where the disease is endemic.^[79]
- Increasingly used to treat scabies (which afflicts around 130 million people worldwide at any one time). Oral ivermectin has been used since 1993 to treat both common scabies and crusted scabies, particularly to control outbreaks in nursing homes where whole-body application of

- topical agents is impractical.^[80] Recently, topical ivermectin lotions were approved, and ivermectin is promising to become the future drug of choice for treating scabies.^[81]
3. The drug of choice to treat difficult-to-treat *Pediculosis capitis* (head lice infestation), the most common parasitic condition among children worldwide.^[82] Oral ivermectin has high efficacy and tolerability, and is more effective than topical malathion lotion.^[83–87] Topical application is also effective.^[88]
 4. An option for ascariasis. Although ivermectin is not recommended for human soil transmitted helminth treatment, except for strongyloidiasis, it has activity against ascariasis, hookworm, and trichuriasis. Relatively few trials have examined the use of ivermectin in this respect. A study to compare the three drugs found that ivermectin was as good as albendazole against ascariasis, but that combination therapy provided slightly better results.^[89] Another study looked at single-dose ivermectin and found it to be as good as three-day albendazole treatment.^[90] Currently, concern is growing about increasing resistance to albendazole and other anthelmintics,^[91] emphasising the need for new control tools.^[92]
 5. The best option for gnathostomiasis. Albendazole and ivermectin are the preferred treatments, but ivermectin is more preferable as it can be given in a single dose.^[93]
 6. An option for mansoniellosis. Ivermectin is highly effective against *Mansonella streptocerca*, with a single dose causing long-term suppression of microfilariae.^[94] However, it has demonstrated little or no effect against *Mansonella perstans*. Although there is no consensus on the best therapy, the most commonly used drug, DEC, is often ineffective and it is likely that combination therapy will be the best option.^[95]
 7. Used widely “off-label” (e.g. to kill skin mites in salmon farming). Toxicity in a range of nontarget animals has been reported, including mice, chicken, rhesus monkeys, bats, and turtles.^[96–100]

Holistic Health, Welfare, and Socioeconomic Impact

Ivermectin is increasingly being viewed as even more of a “wonder drug” in human health, as it has also been improving the nutrition, general health, and well-being of billions of people worldwide ever since it was first used to treat onchocerciasis in humans in 1988. It is ideal in many ways, being multipurpose, highly effective and broad-spectrum, safe, well-tolerated, and can be easily administered (a single, annual oral dose).

Over the 25-year period that communities in Africa and Latin America have been taking ivermectin to combat river blindness and elephantiasis, anecdotal reports of secondary and nontarget benefits have been burgeoning. The benefits described range from an increase in the libido of men to the ability of the tablets to kill termites. Research is accelerating to explore the veracity of these perceived additional benefits and to try and quantify the true overall impact that ivermectin may provide in communities undergoing MDA.

From a purely medical standpoint, ivermectin is known to kill a range of intestinal parasitic worms. The outcome is visible and tangible: people observing worms in their stools. Consequently, owing to this outward manifestation, villagers feel better and are simultaneously encouraged to continue complying with the drug regime.

Work in Brazil investigating the overall health impact of ivermectin in MDA communities indicates that after two standard doses of ivermectin given 10 days apart, intestinal worm burdens are decimated. Infestations with *Strongyloides*, *Enterobius*, and *Ascaris* were completely cured, whereas other worm burdens were cut to 50–85% of their original levels. With regard to external parasites, 99% of pediculosis was cured, compared with scabies (88%) and tungiasis (64%).^[101,102] Another analysis showed that children in a community that underwent 17 years of ivermectin treatment showed markedly reduced prevalence and intensities of *Trichuris trichiuria* infections, and that even children not eligible for treatment displayed reductions, indicating that ivermectin benefitted all members of the community by helping to reduce transmission.^[103]

In a survey of 3125 community members in Nigeria who had been receiving ivermectin MDA, the results were also diverse and impressive. Among those treated, with regard to onchocerciasis, there was an 18.5% reduction in body itching, along with reduced skin rash (17.3%), reports of 11.7% better vision, and a 6.6% darkening of leopard skin. Moreover, in addition to the targeted improvements, 24.6% of individuals reported being dewormed, 22.3% said their appetite had increased, 7.9% felt that they had experienced a noticeable reduction in arthritic or other musculoskeletal pain, 6.6% of men declared their libido had improved, 4.5% of community members said their head lice had disappeared, and 4.5% of women described a reversal of secondary amenorrhea.^[104]

In a subsequent comprehensive four-country study of MDA patients in Africa, diverse health and social impacts and perceptions were quantified (Figure 28). Overall, 84.7% felt ivermectin had provided multiple and substantial health and social welfare benefits. All patients reported being better able to sleep at night and were of the opinion that the MDA had

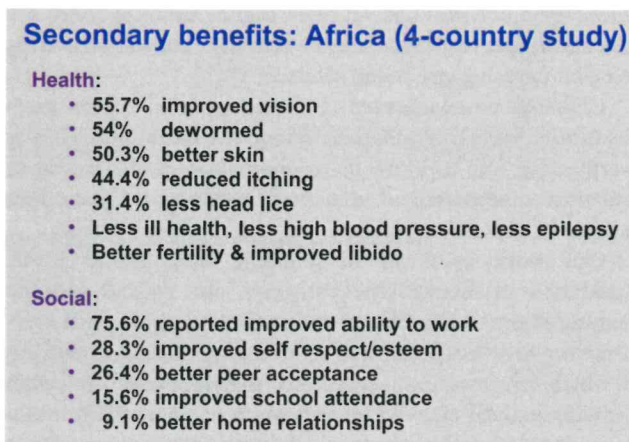


Figure 28. Ivermectin MDA. Source: J. C. Okeibunor et al. 2011.

improved their social, psychological, and economic well-being, with both food productivity and food security being improved.^[105]

Benefits for Japan

The discovery of avermectin has contributed greatly towards improving lives and living standards of billions of people around the world, as well as to improve the health of hundreds of billions of livestock and pets. Development, donation, and distribution of the drug have been associated with many highly beneficial precedents. The substantial royalties earned by the Kitasato Institute on sales of ivermectin in Animal Health have also been used wisely and beneficently. They have funded a great deal of highly focused research, been used to obtain the land covering 27 ha at Kitamoto City in Saitama Prefecture, and to construct a vaccine production facility as well as a 440-bed district general hospital and a nursing college. At present, over 1000 patients per day visit the hospital, which covers a catchment area that was previously grossly underserved with medical facilities. We placed a ceramic plate of a scanning electron micrograph of *S. avermectinius* at the entrance hall of the Kitamoto hospital to illustrate the true foundations on which the building has been constructed and to remind us all of the bounty that still lies hidden in soil, in Japan and elsewhere, awaiting discovery.

Genetics of *S. avermectinius* and Avermectin Biosynthesis.

Soon after its use became widespread in animal health, ivermectin resistance began to appear, at first in small ruminants but also, more significantly in cattle parasites, especially *Cooperia* spp.^[106] It is well-known that high-level resistance to ivermectin appears in free-living *Caenorhabditis elegans*.^[107] Thankfully, despite over 30 years of constant worldwide use, there have been no reports of resistance in canine heartworms or among equine *Strongyloides* parasites. More importantly, despite some 25 years of constant monotherapy in humans, no convincing evidence of resistance in *Onchocerca volvulus* has yet been found, although there are indications that resistance may be starting to develop and that resistant parasites are being selected.^[108,109]

Chemists have achieved the total synthesis of the avermectins. However, to fully understand the biosynthesis of the avermectins, and to allow us to manipulate *S. avermectinius* into producing modified analogues, we mapped the entire genome of the microorganism.

Our work in terms of mapping biosynthetic genes, elucidation of biosynthetic pathways, and overall genome analysis of the avermectin-producing microorganism *S. avermectinius* MA-4680T allowed us to create mutant organisms in which avermectin biosynthesis was blocked. Thorough stepwise analysis allowed identification of single-point mutations, elucidating the structures of biosynthetic intermediates produced by each mutant, and determination of their

locations in the biosynthetic pathways. Moreover, the information taken from these blocked mutants became the basis for the cloning of gene clusters for avermectin biosynthesis.

In 1999, we reported that 17 genes of *S. avermectinius* encode enzymes that are involved in avermectin biosynthesis.^[110–113] Of these, those encoding four types of polyketide synthases I are concerned with lactone formation, via 12 cycles and 53 steps. The remainder act on pathway-specific regulation, with 12 genes being involved in modification of the lactone ring, biosynthesis of oleandrose, and its glycosylation.

The functions of the 17 genes were analyzed by cloning. As shown in Figure 29, four genes, *aveA1*, *aveA2*, *aveA3*, and *aveA4*, are involved in the biosynthesis of the basic skeleton of the aglycone moiety. AVES1–AVES4, whose synthesis is governed by these four genes, are multifunctional proteins composed of 3973, 6239, 5532, and 4681 amino acids, respectively.

There are a total of 12 modules in these four large, multifunctional proteins. The acyltransferase (AT) domain transports acyl groups necessary for acyl-chain elongation, one after another, to the ACP (acyl carrier protein) domain present in each module. The acyl groups are then condensed by the catalytic action of the β -oxoacyl-ACP synthase (KS) domain. The resultant β -oxoacyl-ACP is reduced by the β -oxoacyl-ACP reductase (KR) domain, and β -hydroxyacyl-ACP is further dehydrated by the dehydratase (DH) domain. The chain-elongation reactions and lactonization at the final step by the thioesterase (TE) domain form the basic skeleton of the lactone, and the nascent lactone is further modified by cytochrome P450 (AveE: CYP171A1) and C5-ketoreductase (AveF) to form avermectin aglycones. Through reaction of the *aveB1*–*aveB7* gene products, namely AveBIIwAveBIII, L-oleandrose is synthesized from glucose-1-phosphate as TDP-L-oleandrose and linked to the aglycone-lactone, thus completing avermectin biosynthesis. The presence of the hydroxy group at position 13, which allows the binding of L-oleandrose, is extremely important, as the presence of two L-oleandroses produces the potent antinematode activity of avermectin. The DH domain in module 7 at AVES3 is originally involved in the C13-OH dehydration reaction, but when histidine is substituted for tyrosine in its catalytic active center (consensus motif: HxxxGxxxxP/S), the domain becomes dysfunctional. Subsequently, biosynthesis progresses, while the hydroxy group at position 13 remains, forming the lactone. This single-point mutation, which has resulted in huge health benefits for humankind, allows the sugar (L-oleandrose) binding and subsequent biosynthesis of avremectin, which has superior anthelmintic activity compared to metabolites without the sugar moiety, such as milbemycin and nemadectin.^[114]

Our group completed the analysis of the entire genome (9025608 bases) of *S. avermectinius* MA-4680T in 2003.^[115,116] The information obtained, which represented the first genome analysis of an industrially important actinomycete, provided a major boost for research into the secondary metabolites of microorganisms. We initially estimated that there were 32 such clusters, finally determining that there are 37 clusters involved (Figure 30).^[117] The

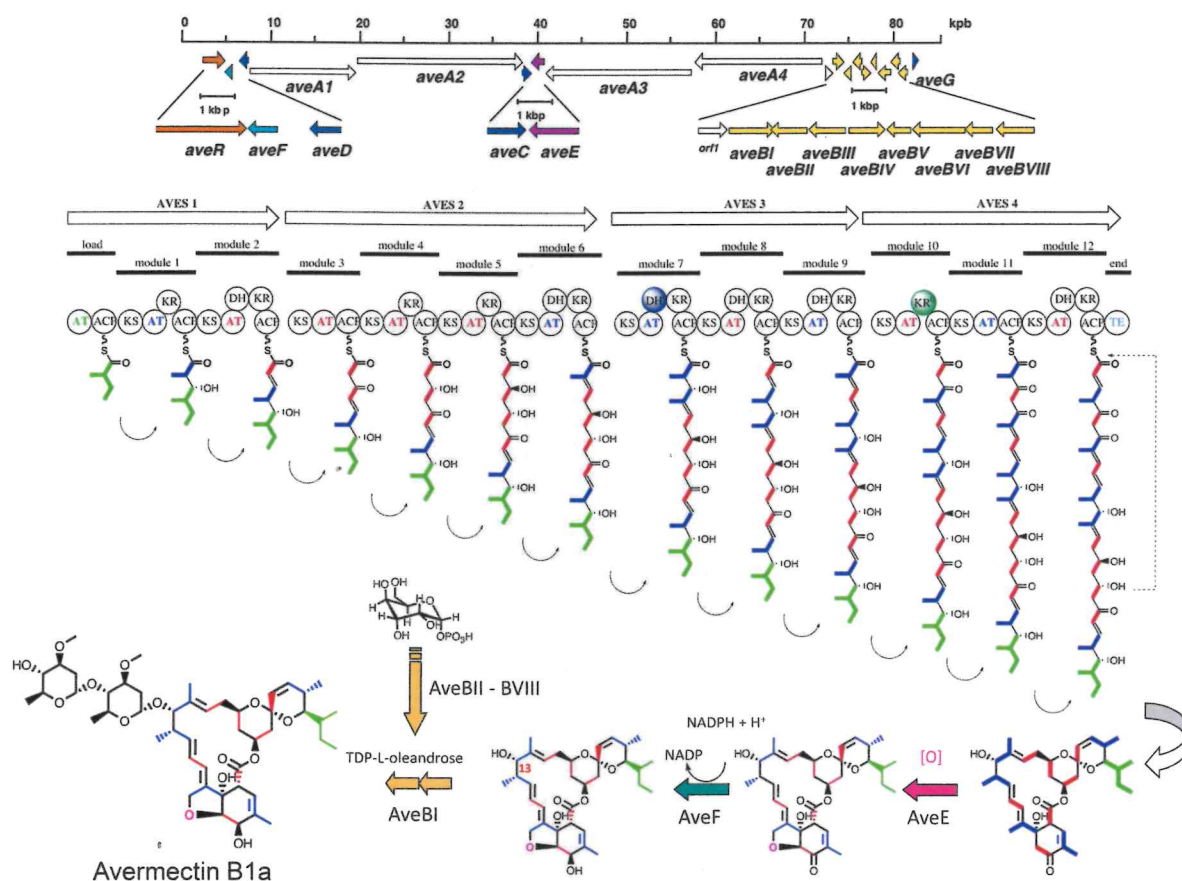
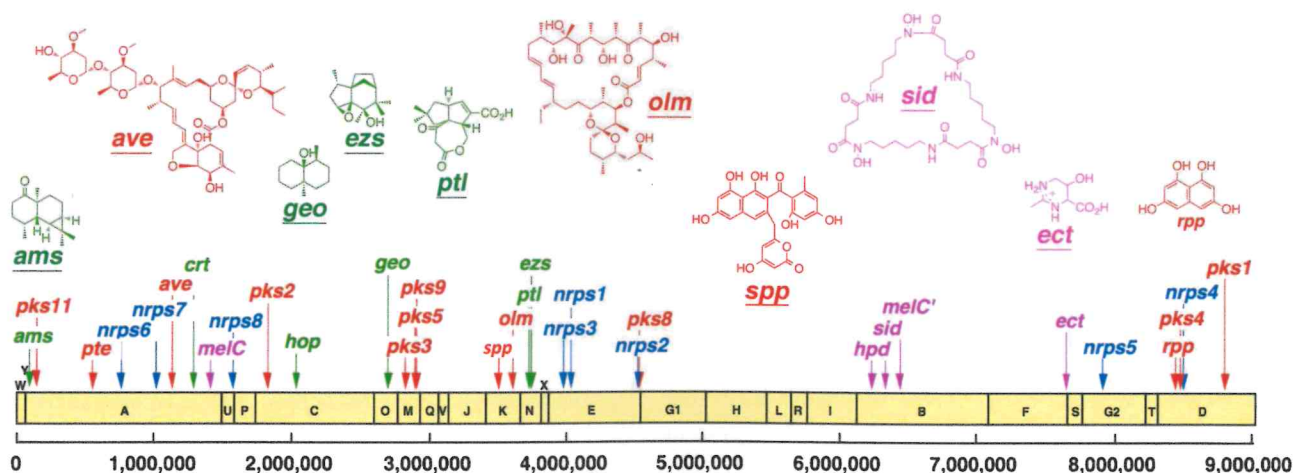


Figure 29. *S. avermectin*: Avermectin biosynthesis.



***Streptomyces avermitilis* (9,025,608 bp; AseI physical map)**

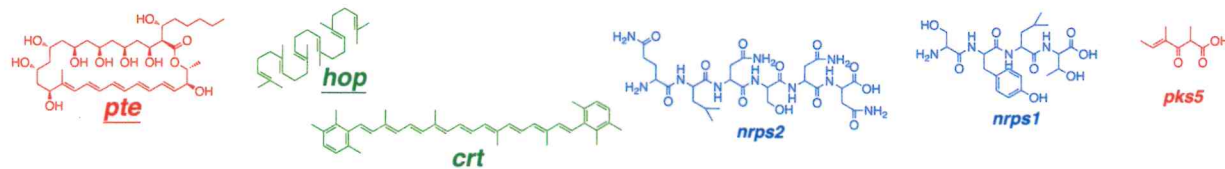


Figure 30. Distribution of gene clusters for secondary metabolite biosyntheses in *Streptomyces avermectin* (*avermitilis*).

production of oligomycin, along with avermectin, was already known, but the production of 10 secondary metabolites, including the polyene macrolide, filipine III (*pte*), carotene (*crt*), pentalenolactone (*ptl*), geosimine (*geo*), and nocardamin (*sid*) were all predicted by the genetic analysis, and later confirmed by isolating each metabolite from a fermentation broth of *S. avermectinius*. This created a new research mechanism, whereby the production of compounds with specific structures can be predicted by gene analysis and later confirmed through actual production and isolation. The mechanism by which secondary metabolites are produced in *S. avermectinius* has now been fully clarified, and work is progressing to engineer the producer microorganism to manufacture yet more potent “designer” compounds.

We have created an improved strain of *S. avermectinius*, which contains only 80% of the original genome, by removing sequences unnecessary for compound production by the site-specific and homologous recombination technique. Using the genome-minimized strain, the heterologous expression of the gene cluster for cephamycin C (Figure 29) biosynthesis from a *S. clavuligerus* genomic library was attempted, resulting in the astonishing production of bioactive compounds.^[118] This is a highly innovative foray in biotechnology, which should provide clues to guide applied research on genetic manipulation and customized culturing systems to facilitate productivity of a range of useful compounds.

Ivermectin: The Future

In addition to the gradual appreciation of the diverse health and socioeconomic benefits that ivermectin does provide, research is beginning to shed light on the promise of ivermectin and the prospects of it combating a range of diseases and for killing vectors of disease-causing parasites. The following list provides an indicator of the potential that has been identified thus far, particularly against diseases of the poor, and provides an insight into the wide spectrum of benefits of ivermectin that may yet lie undiscovered and unexploited.

1. Streptocerciasis: occurs in Central Africa due to infection with the nematode *Dipetalonema streptocerca* transmitted by the bite of insects of the genus *Culicoides*. Ivermectin kills the disease-causing microfilaria.^[119]
2. Trichinosis: globally, 11 million individuals are infected with *Trichinella* roundworms, which can be killed by ivermectin.^[120]
3. Myiasis: infestation by fly larvae that grow inside the host. It is a relatively common affliction of people in poor, rural tropical communities. Surgical removal of the parasites is often the only therapy. Oral myiasis has been successfully treated with ivermectin.^[121]
4. Vector control: The avermectins are toxic to almost all insects, causing water balance difficulties, as well as disruption of moulting and metamorphosis, with death occurring between 1 and 30 days.^[122] Ivermectin kills a wide variety of insects^[123,124] and is highly effective against bedbugs, capable of eradicating or preventing bedbug infestations.^[125]
5. Malaria: Mosquitos (*Anopheles gambiae*) that transmit *Plasmodium falciparum*, the most dangerous malaria-causing parasite in Africa, can be killed by the ivermectin present in the human bloodstream after a standard oral dose.^[126–128] At sub-micromolecular levels, ivermectin inhibits the nuclear import of polypeptides of the signal recognition particle of *P. falciparum* (PfSRP), killing the parasites. This raises the possibility that ivermectin could become a useful, novel malaria-transmission control tool.^[129,130]
6. Leishmaniasis: Ivermectin kills sandflies (*Phlebotomus papatasi*) that transmit the parasites that cause leishmaniasis and has been suggested as a means to help control them.^[131,132] Ivermectin also kills various stages of the disease-causing parasite, *Leishmania major*.^[133,134]
7. Trypanosomiasis: Ivermectin has promise as a systemic drug against the tsetse fly vectors of African trypanosomiasis (sleeping sickness).^[135,136] There is scope for investigating the use of ivermectin for the treatment of trypanosomiasis from several aspects.^[137]
8. Schistosomiasis: A research collaboration was established between the Kitasato Institute and the Oswaldo Cruz Institute (Fiocruz) in Brazil in 2008 to test ivermectin analogues and compounds from the chemical libraries of each institute in screening systems being operated in the two institutions. Promising results were immediately found with regard to the impact of ivermectin on the intermediate host snails responsible for maintaining the schistosomiasis re-infection cycle, offering the prospect of using ivermectin to help control one of the world's major neglected diseases.
9. Antiviral: Ivermectin is a broad-spectrum inhibitor of importin a/b nuclear import, demonstrating potent antiviral activity towards HIV-1 and dengue viruses.^[138] Ivermectin also strongly inhibits replication of several flaviviruses (yellow fever dengue, Japanese encephalitis, and tick-borne encephalitis).^[139,140]
10. Antibacterial: Ivermectin prevents *Chlamydia trachomatis* infection.^[141] It is also reported to be bactericidal against a range of mycobacterial species, including *Mycobacterium tuberculosis*^[142] and *M. ulcerans*.^[143]
11. Anticancer: Ivermectin promotes cell death in leukaemia cells and ME-180 cervical cancer cells.^[144–146]

Concluding Remarks

Ivermectin has continually proved to be astonishingly safe for human use. Indeed, it is such a safe drug, with minimal side effects, that it can be administered by nonmedical staff and even illiterate individuals in remote rural communities, provided that they have had some very basic, appropriate training. This fact has helped contribute to the unsurpassed beneficial impact that the drug has had on human health and welfare around the globe, especially with regard to the campaign to fight onchocerciasis.

In reality, the renewed interest in fighting tropical diseases, including the involvement of the pharmaceutical industry, which has become increasingly evident over the past

four decades, and which has saved lives and improved the welfare of billions of people, notably the poor and disadvantaged in the tropics, can be traced back to the 1987 introduction of ivermectin for use in humans. The remarkable and unparalleled donation of ivermectin can rightly be seen to be the origin of this philanthropic largesse.

Today, ivermectin is being increasingly used worldwide to combat other diseases in humans, and new and promising properties and uses for ivermectin and other avermectin derivatives are continuing to be found. Of perhaps even greater significance is the evidence that the use of ivermectin has both direct and indirect beneficial impact on improving community health. Above all, ivermectin has proved to be a medicine of choice for the world's rural poor.

According to many experts, a post-antibiotic era—in which common infections and minor injuries can kill—is a very real possibility, with WHO Director General Dr. Margaret Chan declaring, “the rise of antibiotic resistance is a global health crisis, and governments now recognize it as one of the greatest challenges for public health today”.

My work has always been guided by five fundamental creeds: 1) the almost unlimited abilities of microorganisms to produce novel compounds; 2) the crucial need to establish “gold-standard” screening systems; 3) recognition that screening is not just a routine exercise; 4) the major contribution of basic research; and 5) the need to apportion highest value to maintaining human relationships and partnerships.

As science advances and our knowledge improves, it is clear to me that the elucidation of suitable targets for medicines, and our expectations for finding remedies to treat both known and as-yet unknown diseases and conditions will not only improve but also accelerate. Genomic mapping and identification of lead compounds have progressed significantly since the turn of the century, as evidenced by the mapping of the human genome. As mentioned above, research is also expected to develop substantially based on the findings of biosynthetic studies and from the investigation of naturally occurring substances that boast hitherto unseen structures. I firmly believe that nature's microbes produce metabolites offering unmatched promise toward meeting our needs, although the introduction of novel screening methods will be key to achieving optimal results. Thus, success will only be restricted by our vision and our innovation—or lack of it. Fortunately, we have access to some of the innovation we need through genetic engineering, and the number of non-natural compounds obtained is increasing rapidly to supplement the never-ending stream of novel compounds that nature can supply.

For 50 years, I have worked alongside specialized researchers in fields such as biochemistry, microbiology, and clinical medicine. My approach has always been influenced by the tenet “one encounter, one chance”. This encompasses the deep reverence that is an essential part of the tea ceremony (or Chanoyu), which is held in the highest esteem in Japanese culture. As well as the certain fact that exact circumstances at any point in time will never happen again, I believe it is important to seize opportunities as and when they arise, and to maintain profound respect and consideration for all my

colleagues—as well as for nature and the microorganisms I work with. Such sentiments form the fundamental basis for all good scientific research and discovery.

Acknowledgements

I would like to convey my sincerest gratitude to all those concerned in my being chosen as a recipient of the 2015 Nobel Prize in Physiology or Medicine, which I humbly receive on behalf of everyone who has helped me at every step of a wonderful journey of discovery. I would also like to express my particular and profound thanks to all the people who have supported me and my research on the discovery, development, and deployment of the avermectins and ivermectin, to Prof. Andy Crump for his invaluable cooperation in the preparation of this account and, of course, to my family for their unwavering and paramount support.

How to cite: *Angew. Chem. Int. Ed.* **2016**, *55*, 10190–10209
Angew. Chem. **2016**, *128*, 10344–10364

- [1] “Mass treatment with ivermectin: an underutilized public health strategy”: R. Speare, D. Durrheim, *Bull. W. H. O.* **2004**, *82*, 559–636.
- [2] “An introduction to the avermectins”: W. C. Campbell, *N. Z. Vet. J.* **1981**, *29*, 174–178.
- [3] “Insecticidal activity of the parasitic avermectins”: D. A. Ostlind, S. Cifelli, R. Lang, *Vet. Rec.* **1979**, *105*, 168.
- [4] “Avermectins: novel insecticides, acaricides and nematocides from a soil microorganism”: I. Putter, J. G. MacConnell, F. A. Preiser, A. A. Haidri, S. S. Ristich, R. A. Dybas, *Experientia* **1981**, *37*, 963–964.
- [5] “Computational improvements reveal great bacterial diversity and high metal toxicity in soil”: J. Gans, M. Wolinsky, J. Dunbar, *Science* **2005**, *309*, 1387–1390.
- [6] “Avermectins, antiparasitic lactones produced by *Streptomyces avermitilis* isolated from a soil in Japan”: E. O. Stapley, H. B. Woodruff, in *Trends in Antibiotic research: Genetics, biosyntheses, actions & new substances* (Eds.: H. Umezawa, A. L. Demain, T. Hata, C. R. Hutchinson), JARA, **1982**, pp. 154–170.
- [7] “Spoxazomicins A–C, novel antitrypanosomal alkaloids produced by an endophytic actinomycete, *Streptosporangium oxazolinicum* K07-0460T”: Y. Inahashi, M. Iwatsuki, A. Ishiyama, M. Namatame, A. Nishihara-Tsukashima, A. Matsumoto, T. Hirose, T. Sunazuka, H. Yamada, K. Otoguro, Y. Takahashi, S. Ōmura, K. Shiomi, *J. Antibiot.* **2011**, *64*, 303–307.
- [8] “Trehangelins A, B and C, novel photo-oxidative hemolysis inhibitors produced by an endophytic actinomycete, *Polymorphospora rubra* K07-0510”: T. Nakashima, R. Okuyama, Y. Kamiya, A. Matsumoto, M. Iwatsuki, Y. Inahashi, K. Yamaji, Y. Takahashi, S. Ōmura, *J. Antibiot.* **2013**, *66*, 311–317.
- [9] “Pyrimidin, a new alkaloid from a *Streptomyces* strain. Taxonomy, fermentation, isolation and biological activity”: S. Ōmura, H. Tanaka, J. Awaya, Y. Narimatsu, Y. Konda, T. Hata, *Agric. Biol. Chem.* **1974**, *38*, 899–906.
- [10] “A new alkaloid AM-2282 of *Streptomyces* origin. Taxonomy, fermentation, isolation and preliminary characterization”: S. Ōmura, Y. Iwai, A. Hirano, A. Nakagawa, J. Awaya, H. Tsuchiya, Y. Takahashi, R. Masuma, *J. Antibiot.* **1977**, *30*, 275–282.

- [11] "Chemical biology of natural indolocarbazole products: 30 years since the discovery of staurosporine": H. Nakano, S. Ōmura, *J. Antibiot.* **2009**, *62*, 17–26.
- [12] "Phosphorylation of Activation Transcription Factor-2 at serine 121 by protein kinase C controls c-Jun-mediated activation of transcription": T. Yamasaki, A. Takahashi, J. Pan, N. Yamaguchi, K. K. Yokoyama, *J. Biol. Chem.* **2009**, *284*, 8567–8581.
- [13] "A novel mode of Gleevec binding is revealed by the structure of spleen tyrosine kinase": S. Atwell, J. M. Adams, J. Badger, M. D. Buchanan, I. K. Feil, K. J. Froning, X. Gao, J. Hendle, K. Keegan, B. C. Leon, H. J. Müller-Dieckmann, V. L. Nienaber, B. W. Noland, K. Post, K. R. Rajashankar, A. Ramos, M. Russell, S. K. Burley, S. G. Buchanan, *J. Biol. Chem.* **2004**, *279*, 55827–55832.
- [14] "Lactacystin, a novel microbial metabolite, induces neurogenesis of neuroblastoma cells": S. Ōmura, T. Fujimoto, K. Otoguro, K. Matsuzaki, R. Moriguchi, H. Tanaka, Y. Sasaki, *J. Antibiot.* **1991**, *44*, 113–116.
- [15] "Structure of prumycin": S. Ōmura, M. Katagiri, K. Atsumi, T. Hata, A. A. Jakubowski, E. Bleecker-Springs, M. Tishler, *J. Chem. Soc. Perkin Trans. 1* **1974**, 1627–1631.
- [16] "Relationships of structure and microbiological activities of the 16-membered macrolides": S. Ōmura, M. Tishler, A. Nakagawa, Y. Hironaka, T. Hata, *J. Med. Chem.* **1972**, *15*, 1011–1015.
- [17] "A new antifungal antibiotic, prumycin": T. Hata, S. Ōmura, M. Katagiri, K. Atsumi, J. Awaya, *J. Antibiot.* **1971**, *24*, 900–901.
- [18] "Studies on cerulenin. III. Isolation and physico-chemical properties of cerulenin": Y. Sano, S. Nomura, Y. Kamio, S. Ōmura, T. Hata, *J. Antibiot.* **1967**, *20*, 344–348.
- [19] "Structure-biological activities relationships among leucomycins and their derivatives": S. Ōmura, M. Katagiri, I. Umezawa, K. Komiyama, T. Maekawa, K. Sekikawa, A. Matsumae, T. Hata, *J. Antibiot.* **1968**, *21*, 532–538.
- [20] "Luminamicin, a new antibiotic. Production, isolation and physico-chemical and biological properties": S. Ōmura, R. Iwata, Y. Iwai, S. Taga, Y. Tanaka, H. Tomoda, *J. Antibiot.* **1985**, *38*, 1322–1326.
- [21] "New antitumor antibiotics, OS-4742 A1, A2, B1 and B2 produced by a strain of *Streptomyces*": S. Ōmura, H. Tanaka, R. Oiwa, J. Awaya, R. Masuma, K. Tanaka, *J. Antibiot.* **1977**, *30*, 908–916.
- [22] "Setamycin, a new antibiotic": S. Ōmura, K. Otoguro, T. Nishikiori, R. Oiwa, Y. Iwai, *J. Antibiot.* **1981**, *34*, 1253–1256.
- [23] "Structure of elasnin, a novel elastase inhibitor": S. Ōmura, A. Nakagawa, H. Ohno, *J. Am. Chem. Soc.* **1979**, *101*, 4386–4388.
- [24] "Factumycin, a new antibiotic (A40A): fermentation, isolation and antibacterial spectrum": V. P. Gullo, S. B. Zimmerman, R. S. Dewey, O. Hensens, P. J. Cassidy, R. Oiwa, S. Ōmura, *J. Antibiot.* **1982**, *35*, 1705–1707.
- [25] "Max Tishler": L. H. Sarrett, C. Roche, in *Biographical memoirs, Vol. 66*, National Academies Press, **1995**, pp. 352–369.
- [26] "Avermectins and related compounds: In Natural Products Isolation: Separation Methods for Antimicrobials": T. W. Miller, V. P. Gullo, *Journal of Chromatography Library, Vol. 43* (Eds.: G. H. Wagman, R. Cooper), Elsevier, Amsterdam, **1989**, pp. 347–376.
- [27] "Ivermectin: a potent new antiparasitic agent": W. C. Campbell, M. H. Fisher, E. O. Stapley, G. Albers-Schonberg, T. A. Jacob, *Science* **1983**, *221*, 823–828.
- [28] "*Streptomyces avermectinius* sp. nov., an avermectin-producing strain": Y. Takahashi, A. Matsumoto, A. Seino, J. Ueno, Y. Iwai, S. Ōmura, *Int. J. Syst. Evol. Microbiol.* **2002**, *52*, 2163–2168.
- [29] "Avermectins: Structure determination": G. Albers-Schonberg, B. H. Arison, J. C. Chabala, A. W. Douglas, P. Eskola, M. H. Fisher, A. Lusi, H. Mrozik, J. L. Smith, R. L. Tolman, *J. Am. Chem. Soc.* **1981**, *103*, 4216–4221.
- [30] "Avermectins, new family of potent anthelmintic agents: producing organisms and fermentation": R. W. Burg, B. M. Miller, E. E. Baker, J. Birnbaum, S. A. Currie, R. Hartman, Y. L. Kong, R. L. Monaghan, G. Olson, I. Putter, J. B. Tunac, H. Wallick, E. O. Stapley, R. Oiwa, S. Ōmura, *Antimicrob. Agents Chemother.* **1979**, *15*, 361–367.
- [31] "Avermectins, new family of potent anthelmintic agents: isolation and chromatographic properties": T. W. Miller, L. Chaiet, D. J. Cole, L. J. Cole, J. E. Flor, R. T. Goegleman, V. P. Gullo, H. Joshua, A. J. Kempf, W. R. Krellwitz, R. L. Monaghan, R. E. Ormond, K. E. Wilson, G. Albers-Schonberg, I. Putter, *Antimicrob. Agents Chemother.* **1979**, *15*, 368–371.
- [32] "Avermectins, new family of potent anthelmintic agents: efficacy of the B1A component": J. R. Egerton, D. A. Ostlund, L. S. Blair, C. H. Eary, D. Suhayda, S. Cifelli, R. F. Riek, W. Campbell, *Antimicrob. Agents Chemother.* **1979**, *15*, 372–378.
- [33] "Ivermectin, a new broad-spectrum antiparasitic agent": J. C. Chabala, H. Mrozik, R. L. Tolman, P. Eskola, A. Lusi, L. H. Peterson, M. F. Woods, M. H. Fisher, W. C. Campbell, *J. Med. Chem.* **1980**, *23*, 1134–1136.
- [34] "Efficacy of the avermectins against filarial parasites: a short review": W. C. Campbell, *Vet. Res. Commun.* **1981**, *5*, 251–262.
- [35] "The avermectins: A new family of antiparasitic agents": I. K. Hotson, *J. South African Vet. Assoc.* **1982**, *53*, 87–90.
- [36] "Investigations of the efficacy of ivermectin against ectoparasites in cattle": D. Barth, I. H. Sutherland, *Zentral. Bakt. Parasit. Infect. Hyg.* **1980**, *57*, 319.
- [37] "On the efficacy of ivermectin versus ticks (*O. moubata*, *R. appendiculatus*, *A. variegatum*) in cattle": C. Centurion, D. Barth, *Zentral. Bakt. Parasit. Infect. Hyg.* **1980**, *58*, 319–320.
- [38] *Ivermectin and Abamectin* (Ed.: W. C. Campbell), Springer, New York, **1989**.
- [39] "A partnership for ivermectin: Social worlds and boundary objects": L. Frost, M. R. Reich, T. Fujisaki, in *Public-Private: Partnerships for Global Health* (Ed.: M. R. Reich), Harvard University Press, **2002**, pp. 87–114.
- [40] S. Ōmura, *Macrolide Antibiotics. Chemistry, Biology and Practice*, 2nd edn (Ed.: S. Ōmura) Academic Press, San Diego, **2002**, pp. 571–576.
- [41] "Actions of dihydroavermectin B1a on insect muscle": R. Duce, R. H. Scott, *Br. J. Pharmacol.* **1985**, *85*, 395–401.
- [42] "*Haemonchus contortus*: ivermectin induced paralysis of the pharynx": T. G. Geary, S. M. Sims, E. M. Thomas, L. Vanover, J. P. Davis, C. A. Winterrowd, R. D. Klein, H. F. Ho, D. P. Thompson, *Exp. Parasitol.* **1993**, *77*, 88–96.
- [43] "Avermectin B1a irreversibly blocks postsynaptic potentials at the lobster neuromuscular junction by reducing muscle membrane resistance": L. C. Fritz, C. C. Wang, A. Gorio, *Proc. Natl Acad. Sci. USA* **1979**, *76*, 2062–2066.
- [44] "Post-synaptic inhibition of invertebrate neuromuscular transmission by avermectin B1a": T. N. Mellin, R. D. Busch, C. C. Wang, *Neuropharmacology* **1983**, *22*, 89–96.
- [45] M. Turner, J. M. Schaeffer, in *Ivermectin and Abamectin* (Ed.: W. C. Campbell), Springer, New York, **1989**, pp. 73–88.
- [46] "Ivermectin: an update": W. C. Campbell, *Parasitol. Today* **1985**, *1*, 10–16.
- [47] "Treatment of 18 children with scabies or cutaneous larva migrans using ivermectin": M. del Mar Saez-De-Ocariz, C. D. McKinster, L. Orozco-Covarrubias, L. Tamayo-Sánchez, R. Ruiz-Maldonado, *Clin. Exp. Dermatol.* **2002**, *27*, 264–267.
- [48] "Immunological tolerance: the key feature in human filariasis?": R. M. Maizels, R. A. Lawrence, *Parasitol. Today* **1991**, *7*, 271–276.

- [49] "The status of ivermectin in the treatment of human onchocerciasis": H. R. Taylor, B. M. Greene, *Am. J. Trop. Med. Hyg.* **1989**, *41*, 460–466.
- [50] "Irreversible effects of ivermectin on adult parasites in onchocerciasis patients in the Onchocerciasis Control Programme in West Africa": A. P. Plaisier, E. S. Alley, B. A. Boatman, G. J. Van Oortmarssen, H. Remme, S. J. De Vlas, L. Bonneux, J. D. Habbema, *J. Infect. Dis.* **1995**, *172*, 204–210.
- [51] "Effect of single-dose ivermectin on *Onchocerca volvulus*: a systematic review and meta-analysis": M.-G. Basáñez, S. D. Pion, E. Boakes, J. A. Filipe, T. S. Churcher, M. Boussinesq, *Lancet Infect. Dis.* **2008**, *8*, 310–322.
- [52] "Migration and death of skin-dwelling *Onchocerca volvulus* microfilariae after treatment with ivermectin": B. O. Duke, G. Soula, G. Zea-Flores, G. L. Bratthauer, O. Doumbo, *Trop. Med. Parasitol.* **1991**, *42*, 25–30.
- [53] "Possible pathogenic pathways in the adverse clinical events seen following ivermectin administrations in onchocerciasis patients": C. D. Mackenzie, T. G. Geary, J. A. Gerlach, *Filaria J.* **2003**, *2*, S5.
- [54] "Ivermectin disrupts the function of the excretory-secretory apparatus in microfilariae of *Brugia malayi*": Y. Moreno, J. F. Nabhan, J. Solomon, C. D. MacKenzie, T. G. Geary, *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 20120–20125.
- [55] "Ivermectin, "Wonder drug" from Japan: the human use perspective": A. Crump, S. Omura, *Proc. Jpn. Acad. Ser. B* **2011**, *87*, 13–28.
- [56] T. Fujisaki, M. Reich, TDR's contribution to the development of ivermectin for onchocerciasis, TDR, Geneva, (TDR/ER/RD/98.3), **1998**.
- [57] WHO/TDR, Participation of the Pharmaceutical sector (TDR/WP/76.30), **1976**.
- [58] "Public-Private Partnerships: Illustrative examples": A. O. Lucas, in *Public-Private Partnerships for Public Health*. (Ed.: M. Reich), Harvard University Press, Cambridge, **2002**, pp. 19–39.
- [59] "History of avermectin and ivermectin, with notes on the history of other macrocyclic lactone antiparasitic agents": W. C. Campbell, *Curr. Pharm. Biotechnol.* **2012**, *13*, 853–865.
- [60] J. L. Sturchio, *The Decision to Donate Mectizan: Historical Background*, Merck & Co., Inc. Rahway, New Jersey, USA, **1992** (unpublished).
- [61] "Efficacy and tolerance of ivermectin in human onchocerciasis": M. A. Aziz, S. Diallo, I. M. Diop, M. Larivière, M. Porta, *Lancet* **1982**, *2*, 171–173.
- [62] "Treatment of human onchocerciasis with ivermectin": J. P. Coulaud, M. Larivière, M. C. Gervais, P. Gaxotte, A. Aziz, A. M. Deluol, J. Cenac, *Bull. Soc. Pathol. Exot. Ses Fil.* **1983**, *76*, 681–688.
- [63] "Ocular findings in a double-blind study of ivermectin versus diethylcarbamazine versus placebo in the treatment of onchocerciasis": K. Y. Dadzie, A. C. Bird, K. Awadzi, H. Schulz-Key, H. M. Gilles, M. A. Aziz, *Br. J. Ophthalmol.* **1987**, *71*, 78–85.
- [64] "Double-blind study of ivermectin and diethylcarbamazine in African onchocerciasis patients with ocular involvement": M. Larivière, M. Aziz, D. Weimann, J. Ginoux, P. Gaxotte, P. Vingtain, B. Beauvais, F. Derouin, H. Schulz-Key, D. Basset, C. Sarfati, *Lancet* **1985**, *326*, 174–177.
- [65] "The status of ivermectin in the treatment of human onchocerciasis": H. R. Taylor, B. M. Greene, *Am. J. Trop. Med. Hyg.* **1989**, *41*, 460–466.
- [66] "The chemotherapy of onchocerciasis II: Quantification of the clinical reaction to microfilaricides": K. Awadzi, *Ann. Trop. Med. Parasitol.* **1980**, *74*, 189–197.
- [67] "The chemotherapy of X. onchocerciasis An assessment of four single dose treatment regimes of MK-933 (ivermectin) in human onchocerciasis": K. Awadzi, K. Y. Dadzie, H. Schulz-Key, D. R. Haddock, H. M. Gilles, M. A. Aziz, *Ann. Trop. Med. Parasitol.* **1985**, *79*, 63–78.
- [68] "Ivermectin as an antiparasitic agent for use in humans": W. C. Campbell, *Annu. Rev. Microbiol.* **1991**, *45*, 445–474.
- [69] "African Programme for Onchocerciasis Control 1995–2015: model-estimated health impact and cost": L. E. Coffeng, W. A. Stolk, H. G. Zouré, J. L. Veerman, K. B. Agblewou, M. E. Murdoch, M. Noma, G. Fobi, J. H. Richardus, D. A. Bundy, D. Habbema, S. J. de Vlas, U. V. Amazigo, *PLoS Neglected Trop. Dis.* **2013**, *7*, e2032.
- [70] *UNESCO World Science Report 2005*, UNESCO, Paris, **2005**, p. 198.
- [71] "Dose-ranging study of ivermectin in the treatment of Filariasis due to *Wuchereria bancrofti*": S. Diallo, M. A. Aziz, O. Ndir, S. Badiane, I. B. Bah, O. Gaye, *Lancet* **1987**, *329*, 1030.
- [72] WHO/TDR, *Tropical Disease Research: Progress 1975–1994*, WHO, Geneva, **1995**, p. 95.
- [73] "Ivermectin for treatment of *Wuchereria bancrofti* filariasis: efficacy and adverse reactions": V. Kumaraswami, E. A. Ottesen, V. Vijayasekaran, *JAMA J. Am. Med. Assoc.* **1988**, *259*, 3150–3153.
- [74] "A controlled trial of ivermectin and diethylcarbamazine in lymphatic filariasis": E. A. Ottesen, V. Kumaraswami, V. Vijayasekaran, *N. Engl. J. Med.* **1990**, *322*, 1113–1117.
- [75] "Comparison of high-dose ivermectin and diethylcarbamazine for activity against Bancroftian filariasis in Haiti": F. O. Richards Jr, M. L. Eberhard, R. T. Bryan, D. F. Mcneeley, P. J. Lammie, M. B. Mcneeley, Y. Bernard, A. W. Hightower, H. C. Spencer, *Am. J. Trop. Med. Hyg.* **1991**, *44*, 3–10.
- [76] Mectizan Donation Program, *Annual Highlights (2104) MDP*, Atlanta, 2015.
- [77] WHO, *Weekly Epidemiological Report*, 90, No. 49, **2015**, pp. 661–680.
- [78] "Ivermectin: panacea for resource-poor communities?": S. Omura, A. Crump, *Trends Parasitol.* **2014**, *30*, 445–455.
- [79] World Health Organization, Strongyloidiasis: Key Facts, World Health Organization, Geneva, **2014**.
- [80] "Oral ivermectin treatment in two cases of scabies: effective in crusted scabies induced by corticosteroid but ineffective in nail scabies": N. Ohtaki, H. Taniguchi, H. Ohtomo, *J. Dermatol.* **2003**, *30*, 411–416.
- [81] "Treatment of scabies: newer perspectives": K. Karthikeyan, *Postgrad. Med. J.* **2005**, *81*, 7–11.
- [82] "Household-wide ivermectin treatment for head lice in an impoverished community: randomized observer-blinded trial": D. Pilger, J. Heukelbach, A. Khakban, F. A. Oliveira, G. Fengler, H. Feldmeier, *Bull. W. H. O.* **2010**, *88*, 90–96.
- [83] "Epidemiology and morbidity of scabies and *Pediculosis capitis* in resource-poor communities in Brazil": J. Heukelbach, T. Wilcke, B. Winter, H. Feldmeier, *Br. J. Dermatol.* **2005**, *153*, 150–156.
- [84] "Oral ivermectin versus malathion lotion for difficult-to-treat head lice": O. Chosidow, B. Giraudeau, J. Cottrell, A. Izri, R. Hofmann, S. G. Mann, I. Burgess, *N. Engl. J. Med.* **2010**, *362*, 896–905.
- [85] "Oral ivermectin for head lice: a comparison with 0.5% topical malathion lotion": A. Nofal, *Z. Hautkrankheiten* **2010**, *8*, 985–988.
- [86] "A pilot study of the use of oral ivermectin to treat head lice in primary school students in Australia": M. J. Currie, G. J. Reynolds, N. Glasgow, *Pediatr. Dermatol.* **2010**, *27*, 595–599.
- [87] "Oral ivermectin for treatment of *Pediculosis capitis*": M. Ameen, R. Arenas, J. Villanueva-Reyes, J. Ruiz-Esmenjaud, D. Millar, F. Domínguez-Dueñas, A. Haddad-Angulo, M. Rodríguez-Alvarez, *Pediatr. Infect. Dis. J.* **2010**, *29*, 991–993.

- [88] "Topical 0.5% ivermectin lotion for treatment of head lice": D. M. Pariser, T. L. Meinking, M. Bell, W. G. Ryan, *N. Engl. J. Med.* **2012**, *367*, 1687–1693.
- [89] "A comparison of the efficacy of single doses of albendazole, ivermectin, and diethylcarbamazine alone or in combinations against *Ascaris* and *Trichuris* spp": V. Y. Belizario, M. E. Amarillo, W. U. de Leon, A. E. de los Reyes, M. G. Bugayong, B. J. Macatangay, *Bull. W. H. O.* **2003**, *81*, 35–42.
- [90] "A comparative trial of a single-dose ivermectin versus three days of albendazole for treatment of *Strongyloides stercoralis* and other soil-transmitted helminth infections in children": H. Marti, H. J. Haji, L. Savioli, H. M. Chwaya, A. F. Mgeni, J. S. Ameir, C. Hatz, *Am. J. Trop. Med. Hyg.* **1996**, *55*, 477–481.
- [91] "School-based health education targeting intestinal worms—further support for integrated control": F. A. Bieri, Y. S. Li, L. P. Yuan, Y. K. He, D. J. Gray, G. M. Williams, D. P. McManus, *PLoS Neglected Trop. Dis.* **2014**, *8*, e2621.
- [92] "Soil-transmitted helminth infections: ascariasis, trichuriasis, and hookworm": J. Bethony, S. Brooker, M. Albonico, S. M. Geiger, A. Loukas, D. Diemert, P. J. Hotez, *Lancet* **2006**, *367*, 1521–1532.
- [93] "Comparison of ivermectin and albendazole treatment for gnathostomiasis": P. Nontasut, V. Bussaratid, S. Chullawichit, N. Charoensook, K. Visetsuk, *Southeast Asian J. Trop. Med. Public Health* **2000**, *31*, 374–377.
- [94] "Long-term suppression of *Mansonella streptocerca* microfilariae after treatment with ivermectin": P. Fischer, E. Tukesiga, D. W. Büttner, *J. Infect. Dis.* **1999**, *180*, 1403–1405.
- [95] "Comparison of different anthelmintic drug regimens against *Mansonella perstans* filariasis": E. R. Bregani, A. Rovellini, N. Mbaidoum, M. G. Magnini, *Trans. R. Soc. Trop. Med. Hyg.* **2006**, *100*, 458–463.
- [96] "Ivermectin toxicity in young mice": B. Skopets, R. P. Wilson, J. W. Griffith, C. M. Lang, *Lab. Anim. Sci.* **1996**, *46*, 111–112.
- [97] "Clinical signs of ivermectin toxicity and efficacy of antiepileptic convulsants as antidotes for ivermectin poisoning in epileptic chickens": J. S. Kim, E. C. Crichlow, *Vet. Hum. Toxicol.* **1995**, *37*, 122–126.
- [98] "Ivermectin toxicology in a rhesus macaque": S. A. Iliff-Sizemore, M. R. Partlow, S. T. Kelley, *Vet. Hum. Toxicol.* **1990**, *32*, 530–532.
- [99] "Ivermectin toxicosis after topical administration in dog-faced fruit bats (*Cynopterus brachyotis*): J. H. DeMarco, D. J. Heard, G. J. Fleming, B. A. Lock, T. J. Scase, *J. Zoo Wildl. Med.* **2002**, *33*, 147–150.
- [100] "Toxicity and efficacy of ivermectin in chelonians": J. A. Teare, M. Bush, *J. Am. Vet. Med. Assoc.* **1983**, *183*, 1195–1197.
- [101] "Selective mass treatment with ivermectin to control intestinal helminthiasis and parasitic skin diseases in a severely affected population": J. Heukelbach, B. Winter, T. Wilcke, M. Muehlen, S. Albrecht, F. A. de Oliveira, L. R. Kerr-Pontes, O. Liesenfeld, H. Feldmeier, *Bull. W. H. O.* **2004**, *82*, 563–571.
- [102] "Efficacy of ivermectin in a patient population concomitantly infected with intestinal helminths and ectoparasites": J. Heukelbach, T. Wilcke, B. Winter, F. A. Sales de Oliveira, R. C. Sabóia Moura, G. Harms, O. Liesenfeld, H. Feldmeier, *Arzneim.-Forsch.* **2004**, *54*, 416–421.
- [103] "Impact of long-term treatment with ivermectin on the prevalence and intensity of soil-transmitted helminth infections": A. L. Moncayo, M. Vaca, L. Amorim, A. Rodriguez, S. Erazo, G. Oviedo, I. Quinzo, M. Padilla, M. Chico, R. Lovato, E. Gomez, M. L. Barreto, P. J. Cooper, *PLoS Neglected Trop. Dis.* **2008**, *2*, e293.
- [104] "The varied beneficial effects of ivermectin (Mectizan) treatment, as observed within onchocerciasis foci in southeastern Nigeria": J. C. Anosike, I. N. Dozie, G. I. Ameh, C. N. Ukaga, B. E. Nwoke, C. T. Nzechukwu, O. S. Udujih, D. C. Nwosu, *Ann. Trop. Med. Parasitol.* **2007**, *101*, 593–600.
- [105] "Where would I be without ivermectin? Capturing the benefits of community-directed treatment with ivermectin in Africa": J. C. Okeibunor, M. Amuyunzu-Nyamongo, N. G. Onyeneho, Y. F. Tchounkeu, C. Manianga, A. T. Kabali, S. Leak, *Trop. Med. Int. Health* **2011**, *16*, 608–621.
- [106] "Drug resistance in nematodes of veterinary importance: a status report": R. M. Kaplan, *Trends Parasitol.* **2004**, *20*, 477–481.
- [107] "The genetics of ivermectin resistance in *C. elegans*": J. A. Dent, *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 2674–2679.
- [108] "Drug resistance in veterinary helminths": A. J. Wolstenholme, *Trends Parasitol.* **2004**, *20*, 469–476.
- [109] "Ivermectin resistance in *Onchocerca volvulus*: Toward a genetic basis": S. Lustigman, J. P. McCarter, *PLoS Neglected Trop. Dis.* **2007**, *1*, e76.
- [110] "Cloning of the gene encoding avermectin B 5-O-methyltransferase in avermectin-producing *Streptomyces avermitilis*": H. Ikeda, L.-R. Wang, T. Ohta, J. Inokoshi, S. Ōmura, *Gene* **1998**, *206*, 175–180.
- [111] "Avermectin biosynthesis": H. Ikeda, S. Ōmura, *Chem. Rev.* **1997**, *97*, 2591–2609.
- [112] "Organization of the biosynthetic gene cluster for the polyketide anthelmintic macrolide avermectin in *Streptomyces avermitilis*": H. Ikeda, T. Nonomiya, M. Usami, T. Ohta, S. Ōmura, *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 9509–9514.
- [113] "Organization of biosynthetic gene cluster for avermectin in *Streptomyces avermitilis*: analysis of enzymatic domains in four polyketide synthases": H. Ikeda, T. Nonomiya, S. Ōmura, *J. Ind. Microbiol. Biotechnol.* **2001**, *27*, 170–176.
- [114] "LL-F28249 antibiotic complex: a new family of antiparasitic macrocyclic lactones. Isolation, characterization and structures of LL-F28249 alpha, beta, gamma, lambda": G. T. Carter, J. A. Nietsche, M. R. Hertz, D. R. Williams, M. M. Siegel, G. O. Morton, J. C. James, D. B. Borders, *J. Antibiot.* **1988**, *41*, 519–529.
- [115] "Genome sequence of an industrial microorganism *Streptomyces avermitilis*: deducing the ability of producing secondary metabolites": S. Ōmura, H. Ikeda, J. Ishikawa, A. Hanamoto, C. Takahashi, M. Shinose, Y. Takahashi, H. Horikawa, H. Nakazawa, T. Osonoe, H. Kikuchi, T. Shiba, Y. Sakaki, M. Hattori, *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 12215–12220.
- [116] "Complete genome sequence and comparative analysis of the industrial microorganism *Streptomyces avermitilis*": H. Ikeda, J. Ishikawa, A. Hanamoto, M. Shinose, H. Kikuchi, T. Shiba, Y. Sakaki, M. Hattori, S. Ōmura, *Nat. Biotechnol.* **2003**, *21*, 526–531.
- [117] "Genomic basis for natural product biosynthetic diversity in the actinomycetes": M. Nett, H. Ikeda, B. S. Moore, *Nat. Prod. Rep.* **2009**, *26*, 1362–1384.
- [118] "Genome-minimized *Streptomyces* host for the heterologous expression of secondary metabolism": M. Komatsu, T. Uchiyama, S. Ō, D. E. Cane, H. Ikeda, *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 2646–2651.
- [119] "Treatment of human *Mansonella streptocerca* infection with ivermectin": P. Fischer, J. Bamuhiga, D. W. Büttner, *Trop. Med. Int. Health* **1997**, *2*, 191–199.
- [120] "Therapeutic potential of myrrh and ivermectin against experimental *Trichinella spiralis* infection in mice": M. M. Basyoni, A. A. El-Sabaa, *Korean J. Parasitol.* **2013**, *51*, 297–304.
- [121] "Oral myiasis treated with ivermectin: case report": E. H. Shinohara, M. Z. Martini, H. G. de Oliveira Neto, A. Takahashi, *Braz. Dent. J.* **2004**, *15*, 79–81.
- [122] "Avermectins in insect control and biology: a review": L. Strong, T. A. Brown, *Bull. Entomol. Res.* **1987**, *77*, 357–389.

- [123] "Ivermectin as a systemic insecticide": H. C. Jackson, *Parasitol. Today* **1989**, *5*, 146–156.
- [124] "Mortality and infertility in adult mosquitoes after the ingestion of blood containing ivermectin": R. B. Tesh, H. Guzman, *Am. J. Trop. Med. Hyg.* **1990**, *43*, 229–233.
- [125] "Ivermectin causes *Cimex lectularius* (Bedbug) morbidity and mortality": J. M. Sheele, J. F. Anderson, T. D. Tran, Y. A. Teng, P. A. Byers, B. S. Ravi, D. E. Sonenshine, *J. Emerg. Med.* **2013**, *45*, 433–440.
- [126] "Effect of ivermectin on *Anopheles gambiae* mosquitoes fed on humans; the potential of oral insecticides in malaria control": C. Chaccour, J. Lines, C. J. M. Whitty, *J. Infect. Dis.* **2010**, *202*, 113–116.
- [127] "The effect of oral anthelmintics on the survivorship and re-feeding frequency of anthropophilic mosquito disease vectors": K. C. Kobylinski, K. M. Deus, M. P. Butters, T. Hongyu, M. Gray, I. M. da Silva, M. Sylla, B. D. Foy, *Acta Trop.* **2010**, *116*, 119–126.
- [128] "Ivermectin mass drug administration for humans disrupts malaria parasite transmission in Senegalese villages": K. C. Kobylinski, M. Sylla, P. L. Chapman, M. D. Sarr, B. D. Foy, *Am. J. Trop. Med. Hyg.* **2011**, *85*, 3–5.
- [129] "*Plasmodium falciparum* signal recognition particle components and anti-parasitic effect of ivermectin in blocking nucleocytoplasmic shuttling of SRP": M. Panchal, K. Rawat, G. Kumar, K. M. Kibria, S. Singh, M. Kalamuddin, A. Mohammed, P. Malhotra, R. Tuteja, *Cell Death Dis.* **2014**, *5*, e994.
- [130] "Endectocides for malaria control": B. D. Foy, K. C. Kobylinski, I. M. da Silva, J. L. Rasgon, M. Sylla, *Trends Parasitol.* **2011**, *27*, 423–428.
- [131] "Ivermectin as a rodent feed-through insecticide for control of immature sand flies (Diptera: Psychodidae)": T. M. Mascari, M. A. Mitchell, E. D. Rowton, L. D. Foil, *J. Am. Mosq. Control Assoc.* **2008**, *24*, 323–326.
- [132] "Comparison between the efficacy of ivermectin and other drugs in treatment of cutaneous leishmaniasis": M. A. Kadir, H. S. Aswad, A. M. Al-Samarai, G. A. Al-Mula, *J. Iraqi Vet. Sci.* **2009**, *23*, 175–180.
- [133] "Effects of ivermectin on blood-feeding *Phlebotomus papatasi* and the promastigote stage of *Leishmania major*": H. A. Hanafi, D. E. Szumlas, D. J. Fryauff, S. S. El-Hossary, G. A. Singer, S. G. Osman, N. Watany, B. D. Furman, D. F. Hoel, *Vector Borne Zoonotic Dis.* **2011**, *11*, 43–52.
- [134] "Efficacy of ivermectin on the infectivity of *Leishmania major* promastigotes": K. A. Rasheid, T. A. Morsy, *J. Egypt Soc. Parasitol.* **1998**, *28*, 207–212.
- [135] "Efficacy of systemic administration of ivermectin against tsetse flies": W. Distelmans, F. D'Haeseleer, J. Mortelmans, *Ann. Soc. Belge Med. Trop.* **1983**, *83*, 119–125.
- [136] "Decrease in survival and fecundity of *Glossina palpalis gambiensis vanderplank 1949* (Diptera; Glossinidae) fed on cattle treated with single doses of ivermectin": S. H. Pooda, K. Mouline, T. De Meeüs, Z. Bengaly, P. Solano, *Parasites Vectors* **2013**, *6*, 165.
- [137] "Effect of ivermectin on *Trypanosoma brucei brucei* in experimentally infected mice": U. K. Udensi, A. F. Fagbenro-Beyioku, *J. Vector Borne Dis.* **2012**, *49*, 143–150.
- [138] "Ivermectin is a specific inhibitor of importin α/β -mediated nuclear import able to inhibit replication of HIV-1 and dengue viruses": K. M. Wagstaff, H. Sivakumaran, S. M. Heaton, D. Harrich, D. A. Jans, *Biochem. J.* **2012**, *443*, 851–856.
- [139] "Ivermectin is a potent inhibitor of flavivirus replication specifically targeting NS3 helicase activity: new prospects for an old drug": E. Mastrangelo, M. Pezzullo, T. De Burghgraeve, S. Kaptein, B. Pastorino, K. Dallmeier, X. de Lamballerie, J. Neyts, A. M. Hanson, D. N. Frick, M. Bolognesi, M. Milani, *J. Antimicrob. Chemother.* **2012**, *67*, 1884–1894.
- [140] "Nuclear localization of dengue virus (DENV) 1–4 non-structural protein 5: protection against all 4 DENV serotypes by the inhibitor ivermectin": M. Y. Tay, J. E. Fraser, W. K. Chan, N. J. Moreland, A. P. Rathore, C. Wang, S. G. Vasudevan, D. A. Jans, *Antiviral Res.* **2013**, *99*, 301–306.
- [141] "Ivermectin inhibits growth of *Chlamydia trachomatis* in epithelial cells": M. A. Pettengill, V. W. Lam, I. Ollawa, C. Marques-da-Silva, D. M. Ojcius, *PLoS ONE* **2012**, *7*, e48456.
- [142] "Anthelmintic avermectins kill *Mycobacterium tuberculosis*, including multidrug-resistant clinical strains": L. E. Lim, C. Vilchère, C. Ng, W. R. Jacobs Jr, S. Ramón-García, C. J. Thompson, *Antimicrob. Agents Chemother.* **2013**, *57*, 1040–1046.
- [143] "In-vitro activity of avermectins against *Mycobacterium ulcerans*": T. F. Omansen, J. L. Porter, P. D. Johnson, T. S. van der Werf, Y. Stienstra, T. P. Stinear, *PLoS Neglected Trop. Dis.* **2015**, *9*, e0003549.
- [144] "The antiparasitic agent ivermectin induces chloride-dependent membrane hyperpolarization and cell death in leukemia cells": S. Sharmeen, M. Skrtic, M. A. Sukhai, R. Hurren, M. Gronda, X. Wang, S. B. Fonseca, H. Sun, T. E. Wood, R. Ward, M. D. Minden, R. A. Batey, A. Datti, J. Wrana, S. O. Kelley, A. D. Schimmer, *Blood* **2010**, *116*, 3593–3603.
- [145] "Identification of therapeutic candidates for chronic lymphocytic leukemia from a library of approved drugs": M. Shen, Y. Zhang, N. Saba, C. P. Austin, A. Wiestner, D. S. Auld, *PLoS ONE* **2013**, *8*, e75252.
- [146] "Potentiation of doxorubicin-induced apoptosis of resistant mouse leukaemia cells by ivermectin": S. Furusawa, H. Shibata, H. Nishimura, S. Nemoto, M. Takayanagi, Y. Takayanagi, K.-I. Sasaki, *Pharm. Pharmacol. Commun.* **2010**, *6*, 129–134.

Received: March 2, 2016

Published online: July 20, 2016